

Genetic association of dopamine beta-hydroxylase (*DBH*) rs72393728 and D-amino acid oxidase activator (*DAOA*) rs3918342 polymorphisms with bipolar disorder in northeastern Algeria

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Abstract: Genes involved in neurotransmitter alteration are implicated in the pathophysiology of bipolar disorder (BD), particularly those belonging to dopaminergic and glutamatergic pathways. This study aimed to investigate the putative association between dopamine beta-hydroxylase (*DBH*) 19bp insertion/deletion rs72393728 and D-amino acid oxidase activator (*DAOA*) M23 rs3918342 polymorphisms with bipolar disorder in a population from northeastern Algeria. The case-control research included 95 patients and 148 controls. The salting out method was applied for the extraction of DNA. PCR was performed to genotype *DBH* rs72393728; genotyping of *DAOA* rs3918342 was conducted by PCR-RFLP. The findings demonstrated significant variations in genotype ($P=0.025$) and allele ($P=0.0088$) frequencies of *DBH* between cases and controls. However, there was no significant correlation between the two groups for *DAOA* either in genotype or allele distributions ($P=0.54$, and $P=0.26$, respectively), although a significant association was detected for grandiose delusion and M23 ($P=0.0015$). The *DBH* rs72393728 polymorphism is reported here for the first time to be associated with the risk of bipolar disorder, both in the Algerian population and in the scientific literature. Despite there being no direct association for *DAOA* rs3918342 polymorphism and bipolar disorder, the two polymorphisms showed significant correlations when combined.

Keywords: dopamine, glutamate, *DBH*, *DAOA*, bipolar disorder

INTRODUCTION

Bipolar disorder (BD) is a severe, lifelong psychiatric disorder characterized by the recurrent alternation of manic and depressive states separated by an euthymic period [1,2]. It is mainly classified into two distinct subtypes: type 1, which typically occurs in mania and depression, and type 2, which presents hypomanic and depressive phases [1,3]. The estimated prevalence of type 1 and type 2 BD is 0.6% and 0.4%, respectively [4,5]. About 1% of the general population suffers from BD [6,7].

BD can manifest through various clinical signs that affect emotion, cognition, and behavior [1]. These manifestations may be explained by the involvement of distinct biological pathways in their pathophysiology. Although the exact cause of BD remains unknown, multiple studies have pointed to a strong genetic component in addition to environmental influences [8]. Genetic susceptibility to BD appears to involve several candidate genes, each contributing a minor effect [9]. Many of these participate in functional pathways [10] and are related to neurotransmitter alteration [11]. Two of the most robustly investigated systems for

their potential role in the pathophysiology of BD are the dopamine and glutamate systems [12,13]. Multiple candidate genes are involved in these pathways, among them dopamine beta-hydroxylase (*DBH*) and D-amino acid oxidase activator (*DAOA*).

DBH is a neurotransmitter-synthesizing enzyme that converts dopamine (DA) into noradrenaline (NA), playing a key role in maintaining the NA/DA balance in the brain [14-16]. The *DBH* gene is mapped to chromosome 9q34 and composed of 12 exons spanning about 23 kb [17]. It contains several potential pathogenic variants, one of which is the 19 bp InsDel rs72393728, which is located 4.5 kb upstream of the transcriptional start site [18]. Increased levels of *DBH* plasmatic activity are associated with manic episodes, while decreased activity is related to depressive episodes in BD patients [15]. *DBH*'s plasmatic activity is genetically controlled, and the *DBH* rs72393728 polymorphism was proven to be strongly associated with it. The Del allele was correlated with reduced promoter activity, causing lower *DBH* levels and enzymatic activity in plasma and cerebrospinal fluid (CSF) [18,19]. A genome-wide association study confirmed the implication of the *DBH* gene as a major determinant of *DBH* plasmatic activity by identifying three variants in the *DBH* promoter region, attributing the strongest association to the rs1611115 polymorphism. Additionally, the study highlighted two other loci (*SARDH* and *LOC338797*) as potential contributors to the regulation of *DBH* expression [20].

The *DAOA* protein is comprised of 153 amino acids and is located in various regions of the central nervous system (CNS). It activates D-amino acid oxidase (DAO), an enzyme crucial for synaptic plasticity and neuronal migration in the brain, and responsible for the degradation of D-serine, a potent activator of N-methyl-D-aspartate (NMDA) type glutamate receptors [21]. Glutamate is an excitatory neurotransmitter implicated in several brain processes [22]. Alterations in the glutamate system have been associated with bipolar disorder (BD), as evidenced by elevated plasma glutamate levels observed in patients with BD [23]. *DAOA* is located on chromosome 13q34, contains 7 exons, and spans 29 kb [21]. The *DAOA* rs3918342, a single-nucleotide polymorphism C>T, also called M23, is situated upstream of the 5'UTR [22], an area that has been found to be positively associated with BD

[24]. It was demonstrated that the T allele of the M23 polymorphism was implicated in both the disturbance of the glutamatergic transmission in BD and dopamine turnover [25]. These findings led to the hypothesis of an eventual connection between glutamatergic and dopaminergic systems, highlighted by Andreou et al. [21], who showed that *DAOA* and *DBH* are linked, despite belonging to distinct pathways. Indeed, the DAO enzyme, after activation by *DAOA*, is involved in the conversion of D-DOPA to L-DOPA, which is the direct precursor of DA; this in turn is converted to NA by the *DBH* enzyme.

Multiple studies have investigated genetic susceptibility to psychiatric disorders. According to Turkish research, the rs6271 variant of *DBH* is associated with BD [26]. However, findings of a recent case-control study conducted in eastern Algeria found no association between the *DBH* rs72393928 polymorphism and schizophrenia (SCZ) [27]. In addition, several single nucleotide polymorphisms (SNPs), including rs5907577 in the intragenic region at Xq27.1 and rs2111504 in the *DPY19L3* gene at 19q13.11, have been linked to bipolar disorder (BD) in a genome-wide association study conducted on European and African American populations, which also confirmed the association of the *ankyrin 3* (*ANK3*) region with BD [28].

The *DAOA* gene (also known as *G72*) and the neighboring region containing the *DAO* (also referred to as *G30*) gene have been extensively analyzed for their genetic association with BD and SCZ in multiple studies. The M23 variant of *DAOA* has been substantially linked to both diseases in a British population [29]. According to family-based research, the *DAOA* rs1935062 polymorphism is specifically correlated to BD [30]. In addition, meta-analyses revealed contradictory findings about the genetic association between *DAOA* and BD [31]. Similarly, *DAO* (*G30*) showed conflicting results regarding its association with BD and SCZ; although several studies have connected the *DAO* gene to SCZ, this correlation could not be confirmed in a British sample [29]. Based on a German investigation, only *DAOA* was linked to BD, whereas both *DAOA* and *DAO* have been related to SCZ [32]. These findings suggest that the *DAOA* (*G72*) gene contributes to susceptibility to both SCZ and BD; however, the considerable variability in results across studies indicates that specific polymorphisms may help differentiate between the two disorders [30].

The main purpose of our study was to explore the implication of two specific genetic polymorphisms: the 19bp InsDel in the *DBH* gene and the M23 single-nucleotide polymorphism in the *DAOA* gene, as potential risk factors for susceptibility to BD in a community of northeastern Algeria. These genes were selected due to their involvement in the dopaminergic and glutamatergic pathways, both of which are strongly implicated in the neurobiology of BD. To the best of our knowledge, this is the first study investigating the association of *DBH* and *DAOA* with bipolar pathology in the Algerian population, thereby providing novel insights to psychiatric genetics research in North Africa.

MATERIALS AND METHODS

Ethics statement

All participants signed a written informed consent to be part of this study. The research protocol was approved by the Ethics Committee of the Dr. Benbadis University Hospital Center of Constantine (Reference Number: CE/CHUC/12/12/2024) and was conducted in accordance with the guidelines of the Declaration of Helsinki.

Study population

This study follows a case-control design. The patient group comprised 95 individuals diagnosed with BD, all of whom met the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5) [33] criteria for BD and were evaluated through structured interviews conducted by experienced psychiatrists. The patients were recruited from the Psychiatric Hospital of Constantine and El-Athmania, and several private medical practices in the region of Constantine, according to the following inclusion criteria: confirmed diagnosis of BD, consenting patients, age ≥ 18 years, and the feasibility of the blood sampling. Medical data, including medical history and clinical features of patients, were collected for analysis. The control group, consisting of 148 healthy individuals, was matched to the patient group by age and gender, and recruited from the same geographical area. None had a personal or family history of psychiatric disorders.

DNA extraction

The genomic DNA was extracted from leukocytes of whole blood samples collected in EDTA-containing tubes using the salting-out method [34]. The purity and concentration of DNA samples were assessed using a Nanodrop spectrophotometer and then stored until use at -20°C .

Genotyping of the *DBH* rs72393728 polymorphism

A PCR (polymerase chain reaction) assay was conducted to genotype the polymorphism rs72393728 of the *DBH* gene using the following primer pair: forward: 5'-GCAAAAGTCAGGCACATGCACC-3'; reverse: 5'-CAATAATTTGGCCTCAATCTTGG-3' [18]. The reaction protocol was carried out as described previously [27]. The obtained PCR products were resolved on a 3% agarose gel (NIPPON Genetics EUROPE, Germany), stained with GelRed (Roth, Germany), and visualized using the Gel Doc XR+ Gel Documentation System (Bio-Rad, USA).

Genotyping of the *DAOA* rs3918342 polymorphism

The polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) assay was performed to genotype the polymorphism rs3918342 of the *DAOA* gene employing the following primer pair: forward: 5'-GGAAACCAGAAAGGTGAAA-3'; reverse: 5'-GAATCAGAAAGGAAAAGTGT-3' [22]. The PCR reaction was prepared in a final volume of 25 μL containing 2.5 μL of $1 \times$ PCR buffer, 0.25 μL of each primer (0.2 μM), 1 μL of dNTP mix (0.2 mM), 2 μL of MgCl_2 (1.5 mM), 0.5 U Taq DNA polymerase (Solis-Biodyne, Estonia), and 50 ng of genomic DNA. The thermocycling program included an initial denaturation at 94°C for 5 min, followed by 30 cycles at 94°C for 30 s (denaturation), 30 s at 58°C (annealing), and 30 s at 72°C (elongation), then a final elongation step at 72°C for 7 min. The amplified PCR products were resolved on a 2% agarose gel (NIPPON Genetics EUROPE, Germany), stained with GelRed (Roth, Germany), and visualized under UV light. The successfully amplified fragments obtained were digested with the *BsaAI* restriction enzyme (New England Biolabs,

Table 1. Clinical characteristics of cases and controls

Demographic and clinical features	Cases (N=95)	Controls (N=148)	P
Age (years) Mean (SD)	42.64 (13.14)	43.4 (13.09)	0.67
Age of onset (years) Mean (SD)	26.46 (9.14)	-	
Gender Male N (%) Female N (%)	37 (38.95) 58 (61.05)	58 (39.19) 90 (60.81)	0.97
Subtype of BD Type 1 N (%) Type 2 N (%)	73 (76.84) 22 (23.16)	- -	
Polarity of first episode Depression N (%) Mania or hypomania N (%)	46 (48.42) 46 (48.42)	- -	
Most recurrent episode Depression N (%) Mania or hypomania N (%)	42 (44.21) 50 (52.63)	- -	
Hospitalization Yes N (%) No N (%)	51 (53.68) 44 (46.32)	- -	
Family history of psychiatric disorders Yes N (%) No N (%)	58 (61.05) 37 (38.95)	- -	
Positive Suicidal history Suicidal attempt N (%) Suicidal ideation N (%)	18 (18.95) 35 (36.84)	- -	
Positive Psychotic Symptoms Delusions N (%) Hallucinations N (%)	60 (63.16) 24 (25.26)	- -	
Tobacco Smokers N (%) Nonsmokers N (%) NA N (%)	27 (28.42) 65 (68.42) 3 (3.16)	20 (13.51) 120 (81.08) 8 (5.41)	0.0142
HTA Yes N (%) No N (%) NA N (%)	13 (13.68) 75 (78.95) 7 (7.37)	14 (9.46) 127 (85.81) 7 (4.73)	0.376
Diabetes Yes N (%) No N (%) NA N (%)	13 (13.68) 75 (78.95) 7 (7.37)	6 (4.05) 135 (91.22) 7 (4.73)	0.0138

N – Number of subjects, SD – standard deviation, NA – not available

USA) for genotyping. To validate the RFLP results, randomly selected PCR products were sequenced directly using a BigDye Terminator and an ABI3500xl genetic analyzer (Applied Biosystems, USA).

Statistical analysis

Statistical analyses were performed using the SPSS software version 26.0 (SPSS Inc., Chicago, Illinois, USA). The Hardy-Weinberg equilibrium (HWE) was assessed separately in the patient and control groups using the chi-square goodness-of-fit test. Association analyses between the patient and control groups for qualitative variables were evaluated using the chi-square test. Logistic regression was applied to analyze genotypic frequencies and to compare type 1 and type 2 BD, genetic models, and the clinical variables within patients. Odds ratios (ORs) and their 95% confidence intervals (CIs) were calculated to measure the significance of the associations. The ANOVA test was used for quantitative variables. Statistical significance was defined as a P value of less than 0.05.

RESULTS

We investigated the potential association of *DBH* 19bp InsDel rs72393728 and *DAOA* M23 rs3918342 polymorphisms, which are involved in dopamine and glutamate systems, respectively, with BD in a population of northeastern Algeria.

Demographic and clinical characteristics

A total of 95 BD patients and 148 healthy controls were included (76.84% with type 1 BD and 23.16% with type 2 BD). The mean age was 42.64±13.14 years for the patients and 43.4±13.09 years for the controls. Both groups had a female predominance, with females comprising 61.05% of the patient group and 60.81% of the control group, resulting in an M/F ratio of 0.64 in both groups. No significant differences were observed between the two groups in terms of age (P=0.67) or gender distribution (P=0.97). However, comparison of demographic and clinical features between bipolar patients and healthy subjects revealed significant differences in the prevalence of tobacco use (P=0.0142) and diabetes (P=0.0138) (Table 1).

Table 2. Genotypic and allelic distributions and genetic models of *DBH* rs72393728 and *DAOA* rs3918342 in cases and controls

Polymorphism		Cases N (%)	Controls N (%)	P	OR (95% CI)	P (adjusted)	OR (95% CI) (adjusted)
DBH rs 72393728	Genotype						
	InsIns	19 (20)	52 (35.14)	0.0298	1.00	0.025	1.00
	InsDel	48 (50.53)	66 (44.59)		1.99 (1.04-3.79)		2.00 (1.05-3.81)
	DelDel	28 (29.47)	30 (20.27)		2.55 (1.22-5.33)		2.60 (1.24-5.44)
	Allele						
	Ins	86 (45.26)	170 (57.43)	0.0088	0.61 (0.42-0.88)	-	-
	Del	104 (54.74)	126 (42.57)				
	Genetic Models						
	Dominant			0.0115	1.00 2.17 (1.18-3.97)	0.0095	1.00 2.18 (1.19-4.00)
	InsIns	19 (20)	52 (35.1)				
InsDel+DelDel	76 (80)	96 (64.9)					
Recessive			0.1012	1.00 1.64 (0.90-2.98)	0.096	1.00 1.67 (0.92-3.04)	
InsIns+InsDel	67 (70.5)	118 (79.7)					
DelDel	28 (29.5)	30 (20.3)					
Log-additive	-	-	-	-	0.0095	1.61 (1.12-2.32)	
DAOA rs3918342	Genotype						
	CC	22 (23.16)	44 (29.73)	0.5267	1.00	0.54	1.00
	CT	47 (49.47)	68 (45.95)		1.38 (0.73-2.60)		1.37 (0.73-2.59)
	TT	26 (27.37)	36 (24.32)		1.44 (0.74-2.96)		1.43 (0.70-2.95)
	Allele						
	C	91 (47.89)	156 (52.70)	0.3014	0.82 (0.57-1.19)	-	-
	T	99 (52.11)	140 (47.30)				
	Genetic Models						
	Dominant			0.2620	1.00 1.40 (0.78-2.54)	0.27	1.00 1.39 (0.77-2.52)
	CC	22 (23.2)	44 (29.7)				
CT+TT	73 (76.8)	104 (70.3)					
Recessive			0.5961	1.00 1.17 (0.65-2.10)	0.6	1.00 1.15 (0.65-1.94)	
CC +CT	69 (72.6)	112 (75.7)					
TT	26 (27.4)	36 (24.3)					
Log-additive	-	-	-	-	0.33	1.20 (0.84-1.71)	

N – number of subjects, Ins – insertion, Del – deletion, OR – odds ratio, CI – confidence interval

Genotyping of the *DBH* rs72393728 polymorphism

The genotype frequencies of the *DBH* rs72393728 polymorphism adhered to Hardy-Weinberg equilibrium in both patient and control groups ($P=0.85$ and 0.33 , respectively), indicating no significant deviation from the normal distribution. Supplementary Fig. S1 illustrates the electrophoresis profile of this polymorphism, showing three different genotypes: a single band of 163pb for homozygous subjects carrying the

insertion allele (InsIns) and a single band of 144pb for homozygous individuals with the deletion allele (DelDel). While heterozygous individuals (InsDel) are characterized by both bands (163 and 144bp).

The association analysis between this polymorphism and BD is presented in Table 2. The results revealed significant differences in both genotype and allele distributions between cases and controls. The genotypic frequency analysis indicates a significant association before and after adjusting for gender and age ($P=0.03$ and 0.025 , respectively), where the DelDel

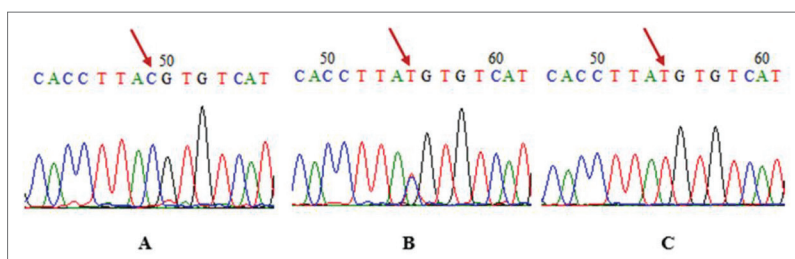


Fig. 1. Direct DNA sequencing results of DAOA rs3918342 polymorphism. **A** – Direct DNA sequencing indicates the presence of 2 copies of the C allele; this patient is homozygote CC. **B** – Direct sequencing indicates the presence of the C allele and the T allele; this patient is heterozygote CT. **C** – Direct DNA sequencing indicates the presence of 2 copies of the T allele; this patient is homozygote TT.

genotype was notably more frequent among cases (29.47%) compared to controls (20.27%), conferring a 2.6-fold increased risk to develop BD compared to the InsIns carriers. Conversely, the frequency of the InsIns genotype was more prevalent in the healthy group (35.14%) than in patients (20%). Allelic frequency analysis also revealed a significant difference between the two groups, with the Del allele being more frequent in cases (54.74%) than in controls (42.57%), supported by a P value of 0.0088.

Results of the genetic model analysis of *DBH* rs72393728 polymorphism are shown in Table 2. In the dominant model, individuals carrying at least one Del allele (InsDel+DelDel) showed a 2.18-fold increased risk of developing BD compared to those with the InsIns genotype, with strong statistical significance ($P=0.0095$). The log-additive model also demonstrated a highly significant association ($P=0.0095$), indicating that each additional copy of the Del allele further increased the risk of BD. In contrast, the recessive model did not reach statistical significance ($P=0.096$), although individuals with the DelDel genotype still showed a 1.61-fold elevated risk.

Genotyping of the DAOA rs3918342 polymorphism

The DAOA rs3918342 polymorphism displayed genotype frequencies consistent with Hardy-Weinberg equilibrium in both patients and controls ($P=0.93$ and 0.39 , respectively), indicating no significant deviation from the expected distribution. Supplementary Fig. S2 presents the electrophoresis profile illustrating the different genotypes of this polymorphism after applying RFLP. Homozygous (CC) individuals present two

bands (356 and 82pb) while homozygous (TT) subjects display a single 440pb band, and heterozygous (CT) individuals are distinguished by the presence of the three bands (440, 356, and 82pb). These results have been confirmed by sequencing for some patients (Fig. 1).

Association analysis results for the DAOA rs3918342 polymorphism (Table 2) globally showed no evidence for any statistically significant differences between patients and healthy participants.

The genotypic frequency analysis indicated that the CC genotype was higher in controls (29.73%) than in cases (23.16%). However, it did not achieve statistical significance. Similarly, the allele frequencies did not differ between the two groups ($P=0.3$). Moreover, when we applied genetic model analyses, no significant association was found under the dominant ($P=0.27$), recessive ($P=0.6$), or log-additive ($P=0.33$) models.

Comparison between type 1 and type 2 BD

Table 3 summarizes the comparison of the association analysis between the two types of BD in patients and controls. Results are presented after adjustment for gender and age. Analysis of *DBH* rs72393728 polymorphism revealed a significant association with type 1 BD ($P=0.015$). Particularly, the DelDel genotype was found to be more frequent among type 1 BD patients (31.5%) than controls (20.3%), with a 3.1-fold higher risk of developing BD. Conversely, the InsIns genotype showed a significantly lower frequency in type 1 BD patients (17.8%) when compared to healthy subjects (35.1%). We did not observe a significant difference between type 2 BD patients and controls ($P=0.79$). Despite the higher frequency of the InsIns genotype that was observed in controls (35.1%) compared to type 2 BD patients (27.3%), there was no statistical significance obtained. *DBH* rs72393728 polymorphism showed a significant association with BD, and more interestingly, it was found to be associated with type 1 BD but not with type 2 BD.

There were notable differences in the genotype distributions of the DAOA rs3918342 polymorphism. The CT genotype was more frequent in controls (46%)

Table 3. Genotypic distribution of *DBH* rs72393728 and *DAOA* rs3918342 in type 1 and type 2 BD

Polymorphism	Genotype	Controls N (%)	BD Type1 N (%)	OR (95% CI)	P Type1 VS Controls	BD Type2 N (%)	OR (95% CI)	P Type2 VS Controls	BD N (%)	OR (95% CI)	P BD VS Controls
<i>DBH</i> rs72393728	InsIns	52 (35.1)	13 (17.8)	1.00	0.015	6 (27.3)	1.00	0.79	19 (20)	1.00	0.025
	InsDel	66 (44.6)	37 (50.7)	2.25 (1.08-4.66)		11 (50)	1.44 (0.550-4.17)		48 (50.5)	2.00 (1.05-3.81)	
	DelDel	30 (20.3)	23 (31.5)	3.10 (1.37-7.01)		5 (22.7)	1.34 (0.37-4.81)		28 (29.5)	2.60 (1.24-5.44)	
<i>DAOA</i> rs3918342	CC	44 (29.7)	14 (19.2)	1.00	0.21	8 (36.4)	1.00	0.24	22 (23.2)	1.00	0.54
	CT	68 (46)	41 (56.2)	1.88 (0.92-3.85)		6 (27.3)	0.48 (0.16-1.49)		47 (49.5)	1.37 (0.73-2.59)	
	TT	36 (24.3)	18 (24.7)	1.61 (0.70-3.70)		8 (36.4)	1.17 (0.40-3.46)		26 (27.4)	1.43 (0.70-2.95)	

N – number of subjects, OR – odds ratio, CI – confidence interval

than in type 2 BD patients (27.3%), while it was more prevalent in type 1 BD patients (56.2%) than in controls (27.3%). Nevertheless, there was no significant association found between patients and healthy individuals in both types of BD ($P=0.21$ for type 1 and $P=0.24$ for type 2).

Association analysis of *DBH* rs72393728 and *DAOA* rs3918342 polymorphisms with clinical variables in patients

Associations between clinical features and the two polymorphisms in the patient group are described in Table 4. Comparison of the genotypic distributions of *DBH* rs72393728 polymorphism with the clinical symptoms of BD did not reveal any substantial associations with most of the clinical variables ($P>0.05$), except for suicidal ideation, which showed a significant association ($P=0.034$). Specifically, the DelDel genotype was notably less frequent among patients with suicidal thoughts (14.3%) compared to those without (38.3%).

Regarding *DAOA* rs3918342 polymorphism, we observed a significant association with gender ($P=0.016$). The CT genotype was more prevalent in males (67.6%) than in females (37.9%), whereas the TT genotype was more frequent in females (34.5%) than in males (16.2%). We also found that grandiose delusions presented a strong significant difference ($P=0.0015$), with the CT genotype being notably more

frequent among patients with this symptom (70%) versus those without (34.5%). For the other clinical variables, no findings of significance were observed.

Association of combined genotypes of *DBH* rs72393728 and *DAOA* rs3918342 polymorphisms with BD

Combined genotype analyses of *DBH* and *DAOA* polymorphisms were performed to assess potential interactions between them. The results shown in Table 5 revealed two important combinations associated with BD. The combination InsIns (*DBH*) – CC (*DAOA*) was significantly more frequent in controls (32.4%) than in BD patients (21.5%), with a P value of 0.0095, indicating a potential protective effect. Conversely, the combination DelDel (*DBH*) and CT (*DAOA*) was more prevalent in patients (39.4%) than in healthy subjects (33.1%), with a P value of 0.0291, showing a significant association with increased risk.

Our case-control study revealed a significant association between *DBH* rs72393728 and BD, with the Del allele as a risk factor. When analyzing each subtype of BD separately, a statistically significant association was detected with only type 1 BD, suggesting a subtype-specific genetic contribution. Additionally, this polymorphism exhibited a significant correlation with suicidal thoughts in BD. However, the *DAOA* rs3918342 polymorphism showed no direct association

Table 4. Association of *DBH* rs72393728 and *DAOA* rs3918342 genotypes with clinical variables in patients

Polymorphism	<i>DBH</i> rs72393728			P	<i>DAOA</i> rs3918342			P
Genotype	InsIns	InsDel	DelDel		CC	CT	TT	
Gender								
Male N (%)	6 (16.2)	20 (54)	11 (29.7)	0.75	6 (16.2)	25 (67.6)	6 (16.2)	0.016
Female N (%)	13 (22.4)	28 (48.3)	17(29.3)		16 (27.6)	22 (37.9)	20 (34.5)	
Age								
Mean (SD)	39.74 (11.03)	43.69 (13.98)	42.82 (13.11)	0.529	48.14 (11.95)	41.47 (13.53)	40.12 (12.53)	0.076
Age of onset								
Mean (SD)	24.44 (7.54)	27.06 (9.66)	26.71 (9.29)	0.713	26.95 (9.83)	26.02 (8.73)	26.8 (9.64)	0.987
Family history of psychotic disorders								
Yes N (%)	11 (19)	34 (58.6)	13 (22.4)	0.1	15 (25.9)	25 (43.1)	18 (31)	0.38
No N (%)	8 (21.6)	14 (37.8)	15 (40.5)		7 (18.9)	22 (59.5)	8 (21.6)	
Subtype of BD								
Type 1 N (%)	13 (17.3)	37 (50.7)	23 (31.5)	0.61	14 (19.2)	41 (56.2)	18 (24.7)	0.072
Type 2 N (%)	6 (27.3)	11 (50)	5 (22.7)		8 (36.4)	6 (27.3)	8 (36.4)	
Polarity of first episode								
Depression N (%)	8 (17.4)	26 (56.5)	12 (26.1)	0.14	9 (19.6)	19 (41.3)	18 (39.1)	0.25
Mania or hypomania N (%)	11 (23.9)	20 (43.5)	15 (32.6)		11 (23.9)	27 (58.7)	8 (17.4)	
Most recurrent episode								
Depression N (%)	8 (19.1)	22 (52.4)	12 (28.6)	0.76	10 (23.8)	18 (42.9)	14 (33.3)	0.83
Mania or hypomania N (%)	11 (22)	24 (48)	15 (30)		10 (20)	28 (56)	12 (24)	
Grandiose delusions								
Yes N (%)	11 (27.5)	21 (52.5)	8 (20)	0.1	7 (17.5)	28 (70)	5 (12.5)	0.0015
No N (%)	8 (14.6)	27 (49.1)	20 (36.4)		15 (27.3)	19 (34.5)	21 (38.2)	
Persecutory delusions								
Yes N (%)	7 (17.9)	20 (51.2)	12 (30.7)	0.91	7 (17.9)	21 (53.8)	11 (28.2)	0.59
No (%)	12 (21.4)	28 (50)	16 (28.6)		15 (26.8)	26 (46.4)	15 (26.8)	
Hallucinations								
Yes N (%)	5 (20.8)	11 (45.8)	8 (33.3)	0.85	3 (12.5)	14 (58.3)	7 (29.2)	0.49
No N (%)	14 (19.7)	37 (52.1)	20 (28.2)		19 (26.8)	33 (46.5)	19 (26.8)	
Suicidal ideation								
Yes N (%)	9 (25.7)	21 (60)	5 (14.3)	0.034	6 (17.1)	20 (57.1)	9 (25.7)	0.55
No N (%)	10 (16.7)	27 (45)	23 (38.3)		16 (26.7)	27 (45)	17 (28.3)	
Suicidal attempt								
Yes N (%)	5 (27.8)	11 (61.1)	2 (11.1)	0.11	4 (22.2)	10 (55.6)	4 (22.2)	0.49
No N (%)	14 (18.2)	37 (48)	26 (33.8)		18 (23.4)	37 (48)	22 (28.6)	
Tobacco								
Smokers N (%)	14 (20.6)	34 (50)	20 (29.4)	0.97	18 (26.5)	29 (42.6)	21 30.9)	0.74
Nonsmokers N (%)	5 (18.5)	15 (51.9)	8 (29.6)		4 (14.8)	18 (66.7)	5 (18.5)	

N – number of subjects, SD – standard deviation

with bipolar pathology, even though it was found to be strongly related to grandiose delusions in BD. On the other hand, the combined genotypes InsIns (*DBH*) and CC (*DAOA*), and DelDel (*DBH*) and CT (*DAOA*) are associated with bipolar illness.

DISCUSSION

BD is a chronic psychiatric illness defined by the presence of recurrent episodes of mania and depression [1]. Being a complex and multifactorial pathology, the implication of genetic factors alongside environmental ones in its etiology is well established [1]. Multiple potential candidate genes were related to neurotransmitter

Table 5. Combined genotypes of *DBH* rs72393728 and *DAOA* rs3918342 in cases and controls

Genotypes	Patients (N)	Patients (frequency)	Controls (N)	Controls (frequency)	X ² (df)	P
InsIns(DBH) CC(DAOA)	41	0.215	96	0.324	6.72(1)	0.0095
InsIns(DBH) CT(DAOA)	66	0.347	120	0.405	1.65(1)	0.1994
InsIns(DBH) TT(DAOA)	45	0.236	88	0.297	2.12(1)	0.1451
InsDel(DBH) CC(DAOA)	70	0.368	110	0.371	0.0051(1)	0.9432
InsDel(DBH) CT(DAOA)	95	0.5	134	0.452	1.04(1)	0.3086
InsDel(DBH) TT(DAOA)	74	0.389	102	0.344	1.0069(1)	0.3156
DelDel(DBH) CC(DAOA)	50	0.263	74	0.25	0.10(1)	0.7457
DelDel(DBH) CT(DAOA)	75	0.394	98	0.331	4.74(1)	0.0291
DelDel(DBH) TT(DAOA)	54	0.284	66	0.222	2.33(1)	0.127

N – number of subjects, X² – chi-square statistic, df – degrees of freedom

alterations [11]. Specifically, dopamine and glutamate neurotransmission have been identified as key factors in the pathophysiology of BD [3,23], and both systems have been extensively studied [12,13]. In the present study, we aimed to assess a potential association of *DBH* rs72393728 and *DAOA* rs3918342 genes, which are members of the dopaminergic and glutamatergic systems, respectively, with BD in an Algerian population.

Various *DBH* polymorphisms have been explored for their association with BD. In this study, we investigated the implication of the 19bp InsDel variant of *DBH* rs72393728. Our results demonstrated a significant association of this polymorphism with BD risk. The Del allele, which was more common among patients, showed a substantial correlation with the pathology. This association suggests that this allele may serve as a susceptibility factor influencing the risk of developing BD in our cohort. Additionally, the genetic models revealed a significant effect of the Del allele under both dominant and log-additive models, indicating that carrying a single copy of this allele increases the risk of BD, with each additional copy further elevating the risk. However, this association was observed only in type 1 BD and not in type 2. To the best of our knowledge, this is the first study to examine the *DBH* 19 bp InsDel variant in the Algerian population and the first to identify an association between this polymorphism and susceptibility to bipolar disorder in the literature. A single study by Zebarjad et al. on

the Iranian population explored the implication of the *DBH* 19 pb InsDel polymorphism rs72393728 in the risk of type 1 BD [6]. In contrast to our findings, their results did not support any association between the studied polymorphism and the risk of type 1 BD; in fact, allele frequencies were similar between cases and unaffected controls [6]. Other *DBH* variants have been examined for their association with BD in British, German, and Turkish populations [35,36,26]; among these, only the Turkish study identified a significant association with rs6271 [26], while no associations were found for the other polymorphisms in the British and German cohorts

[35,36]. BD shares overlapping clinical symptoms with other psychiatric disorders, such as major depressive disorder (MDD) and schizophrenia (SCZ), especially with MDD during depressive episodes [15,16]. Due to these symptomatic similarities, it is hypothesized that these three pathologies may share common genetic predisposition variants [2]. Thus, the involvement of *DBH* has been investigated through association studies not only with BD, but also with MDD and SCZ. An association of *DBH* rs72393728 polymorphism with MDD was detected in Danish and Chinese populations [37,38], highlighting the effect of the Del allele as a risk factor. Nevertheless, no association was registered with SCZ [18,39,40], including a recent study involving the Algerian population [27].

From a neurobiological perspective, the balance between dopamine (DA) and noradrenaline (NA) plays a crucial role in both emotion regulation and cognitive function, mediated by the *DBH* enzyme that catalyzes the conversion of DA to NA [14,15]. The rate of *DBH* protein production is tightly genetically controlled [19]. Indeed, variants of the *DBH* gene, including the 19 pb InsDel, are known to be significantly related to *DBH* enzymatic activity [16,19,41]. Specifically, the Ins allele of *DBH* rs72393728 is associated with higher plasma activity [14], whereas the Del allele is associated with lower plasma activity of *DBH* [41,42]. In fact, the Del allele has been linked to reduced promoter activity, leading to decreased enzymatic activity of *DBH* in the brain, resulting in less efficient conversion of DA to

NA [19]. The disruption in the DA/NA ratio caused by the Del allele effect is involved with depression [37].

Research has shown a strong correlation between dysregulation of DBH enzyme activity and the mood phases of bipolar disorder, with elevated plasma DBH activity observed in manic episodes and reduced activity during depressive episodes [16,23]. It has also been reported that plasmatic variations in DBH activity may be associated with the psychotic symptoms in SCZ [43]. Consequently, an altered enzymatic activity of DBH was related to mental disorders, and a low plasma DBH activity is considered a risk factor [16,37,44]. In this context, the *DBH* gene may serve as a marker for common psychiatric symptoms [19,37].

Herein, the *DBH* rs72393728 polymorphism was found to be associated with suicidal ideations in BD patients. Indeed, we registered that patients who reported suicidal thoughts differed significantly from those who did not in terms of their genotype. However, suicidal attempts did not show any statistical relevance. Very few studies have considered analyzing the relationship between the *DBH* gene and suicidal behavior in BD and other psychiatric disorders. One study examined the 19 bp InsDel polymorphism of *DBH* rs72393728 in relation to suicide attempts among schizophrenic patients. The findings reported no correlation [42].

The *DAOA* gene has elicited considerable scientific interest, not only due to its implication in the glutamate system, which has a crucial role in excitatory neurotransmission in the brain [13,23], but also due to its location on chromosome 13q34, a region strongly associated with psychiatric disorders (13q34), particularly BD and SCZ [23,39]. The *DAOA* rs3918342 polymorphism is one of the most commonly investigated for its involvement in the pathophysiology of BD [46]. However, results are conflicting across studies [22,46].

Our results showed no significant association between the *DAOA* M23 polymorphism and BD, as neither genotype nor allele distributions differed significantly between patients and healthy controls. The same results were obtained when applying genetic models (all *P* values >0.05). Consistent with our findings, two studies conducted in Chinese populations [5,46], as well as studies in Canadian [47] and German [48] populations, reported no association between the *DAOA* (rs3918342) polymorphism and BD, suggesting

that this variant does not contribute to susceptibility to the illness. Conversely, the results of other studies carried out on British [29] and Iranian [22] populations demonstrated the presence of a significant association of M23 with BD, with the T allele being frequently more common in patients than in controls [22,29], suggesting that it may be a risk factor for susceptibility to BD.

In another exploratory study of the German population [45], M23 polymorphism was related to BD patients with psychotic symptoms, particularly those with persecutory delusions. The C allele was more prevalent in these patients, while patients with no psychotic signs showed a similar genotypic distribution as the control group. In the same study conducted in a Polish population, identical results were obtained [45].

In addition to its individual role, *DAOA* has also been studied in combination with the adjacent *G30* gene in haplotype analyses for their association with BD and SCZ [49,50]. *DAOA*/*G30* haplotypes have been reported to be associated with psychotic features and mood alterations involving delusions in Romanian patients with BD [51,52], suggesting that these haplotypes may influence affective psychosis without directly contributing to the pathogenesis of BD [52]. Our findings align with these results regarding M23 and psychotic features. In our cohort, the association involved the subgroup with grandiose delusions compared to patients without this psychotic symptom. Accordingly, based on the findings from our study and the aforementioned research, the *DAOA* rs3918342 polymorphism may not be directly associated with susceptibility to developing bipolar disorder. However, it may be linked to psychotic features of the illness.

Neurobiological evidence may offer a clearer understanding of the involvement of the *DAOA* gene and the glutamatergic system in the pathophysiology of bipolar disorder. Glutamate signaling dysfunction plays a critical role in the etiopathogenesis of BD [10], as reflected by elevated glutamate levels observed in both the plasma of BD patients [23] and in post-mortem brain tissue samples [23,53]. A meta-analysis also reported that brain levels of glutamate and glutamine were increased in bipolar patients [54]. Alves and colleagues, in their review, reported that elevated glutamatergic function was related to depression [52]. Imaging genetics approaches have potentially related

genetics to structural brain phenotypes. Indeed, alterations in the *DAOA* gene have been associated with reductions in gray matter volume [11]. Another study conducted by Zuliani et al. revealed that M23 polymorphism of the *DAOA* gene may influence structural brain abnormality, where the TT genotype was found to be associated with temporal pole and amygdala gray matter reduction in BD patients [55].

A possible interaction between *DBH* and *DAOA* genes was revealed in our findings of combined genotypes. The genotype InsIns (*DBH*) and CC (*DAOA*) was more frequent among unaffected subjects, which indicates a potential protective effect against BD for individuals carrying this combination, while the DelDel (*DBH*) and CT (*DAOA*) genotypes were more common in patients, suggesting its involvement in BD susceptibility as a risk factor. It has been demonstrated that psychosis may be caused by an interaction between dopamine and glutamate systems, where increased dopamine function may be linked to glutamatergic dysfunction in psychotic disorders [56]. These findings have substantiated this hypothesis, although further research on this interaction is necessary to elucidate the mechanism of action between these two systems.

Our study had certain limitations that should be acknowledged. The primary limitation was the relatively small sample size, which, although sufficient to detect meaningful associations, may increase the risk of both false-positive and false-negative results. Secondly, the scarcity of prior research on *DBH* and BD limited the comparison of our results with existing data. Therefore, to corroborate the observed findings, the reproducibility of the results must be validated in further research involving larger samples and diverse demographics. It is also recommended to expand the genetic analysis by investigating a wider range of polymorphisms within the dopaminergic and glutamatergic pathways.

In conclusion, this study provides novel insights into the involvement of the *DBH* rs72393728 polymorphism in bipolar disorder, identifying the Del allele as a risk factor for susceptibility in the northeastern Algerian population. Conversely, the *DAOA* rs3918342 polymorphism did not show a direct association with BD, while findings revealed a significant association with grandiose delusions in our cohort. Nonetheless,

our results suggest a potential interaction between these two polymorphisms. This study highlights the genetic diversity of bipolar disorder and the distinct pathways that may contribute to its etiology, underscoring the need for further research to elucidate the precise mechanisms underlying these associations.

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REFERENCES

1. Stahl EA, Breen G, Forstner AJ, McQuillin A, Ripke S, Trubetskoy V, Mattheisen M, Wang Y, Coleman JRI, Gaspar HA, de Leeuw CA, Steinberg S, Whitehead Pavlides JM, Trzaskowski M, Byrne EM, Pers TH, Holmans PA, Richards AL, Abbott L, Agerbo E, Akil H, Albani D, Alliey-Rodriguez N, Als TD, Anjorin A, Antilla V, Awasthi S, Badner JA, Bækvad-Hansen M, Barchas JD, Bass N, Bauer M, Belliveau R, Bergen SE, Bøcker Pedersen C, Bøen E, Boks MP, Boocock J, Budde M, Bunney W, Burmeister M, Bybjerg-Grauholm J, Byerley W, Casas M, Cerrato F, Cervantes P, Chambert K, Charney AW, Chen D, Churchhouse C, Clarke TK, Coryell W, Craig DW, Cruceanu C, Curtis D, Czerski PM, Dale AM, de Jong S, Degenhardt F, Del-Favero J, DePaulo JR, Djurovic S, Dobbyn AL, Dumont A, Elvsåshagen T, Escott-Price V, Fan CC, Fischer SB, Flickinger M, Foroud TM, Forty L, Frank J, Fraser C, Freimer NB, Frisén L, Gade K, Gage D, Garnham J, Giambartolomei C, Giørtz Pedersen M, Goldstein J, Gordon SD, Gordon-Smith K, Green EK, Green MJ, Greenwood TA, Grove J, Guan W, Guzman-Parra J, Hamshere ML, Hautzinger M, Heilbronner U, Herms S, Hipolito M, Hoffmann P, Holland D, Huckins L, Jamain S, Johnson JS, Juréus A, Kandaswamy R, Karlsson R, Kennedy JL, Kittel-Schneider S, Knowles JA, Kogevinas M, Koller AC, Kupka R, Lavebratt C, Lawrence J, Lawson WB, Leber M, Lee PH, Levy SE, Li

- JZ, Liu C, Lucae S, Maaser A, MacIntyre DJ, Mahon PB, Maier W, Martinsson L, McCarroll S, McGuffin P, McInnis MG, McKay JD, Medeiros H, Medland SE, Meng F, Milani L, Montgomery GW, Morris DW, Mühleisen TW, Mullins N, Nguyen H, Nievergelt CM, Nordin Adolfsson A, Nwulia EA, O'Donovan C, Olde Loohuis LM, Ori APS, Oruc L, Ösby U, Perlis RH, Perry A, Pfennig A, Potash JB, Purcell SM, Regeer EJ, Reif A, Reinbold CS, Rice JP, Rivas F, Rivera M, Roussos P, Ruderfer DM, Ryu E, Sánchez-Mora C, Schatzberg AF, Scheftner WA, Schork NJ, Shannon Weickert C, Shekhtman T, Shilling PD, Sigurdsson E, Slaney C, Smeland OB, Sobell JL, Søholm Hansen C, Spijker AT, St Clair D, Steffens M, Strauss JS, Streit F, Strohmaier J, Szelinger S, Thompson RC, Thorgeirsson TE, Treutlein J, Vedder H, Wang W, Watson SJ, Weickert TW, Witt SH, Xi S, Xu W, Young AH, Zandi P, Zhang P, Zöllner S, eQTLGen Consortium, BIOS Consortium, Adolfsson R, Agartz I, Alda M, Backlund L, Baune BT, Bellivier F, Berrettini WH, Biernacka JM, Blackwood DHR, Boehnke M, Børglum AD, Corvin A, Craddock N, Daly MJ, Dannlowski U, Esko T, Etain B, Frye M, Fullerton JM, Gershon ES, Gill M, Goes F, Grigoriou-Serbanescu M, Hauser J, Hougaard DM, Hultman CM, Jones I, Jones LA, Kahn RS, Kirov G, Landén M, Leboyer M, Lewis CM, Li QS, Lissowska J, Martin NG, Mayoral F, McElroy SL, McIntosh AM, McMahon FJ, Melle I, Metspalu A, Mitchell PB, Morken G, Mors O, Mortensen PB, Müller-Myhsok B, Myers RM, Neale BM, Nimgaonkar V, Nordentoft M, Nöthen MM, O'Donovan MC, Oedegaard KJ, Owen MJ, Paciga SA, Pato C, Pato MT, Posthuma D, Ramos-Quiroga JA, Ribasés M, Rietschel M, Rouleau GA, Schalling M, Schofield PR, Schulze TG, Serretti A, Smoller JW, Stefansson H, Stefansson K, Stordal E, Sullivan PF, Turecki G, Vaaler AE, Vieta E, Vincent JB, Werge T, Nurnberger JI, Wray NR, Di Florio A, Edenberg HJ, Cichon S, Ophoff RA, Scott LJ, Andreasen OA, Kelsoe J, Sklar P, Bipolar Disorder Working Group of the Psychiatric Genomics Consortium. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nat Genet.* 2019;51(5):793-803.
<https://doi.org/10.1038/s41588-019-0397-8>
2. Rolstad S, Pålsson E, Ekman CJ, Eriksson E, Sellgren C, Landén M. Polymorphisms of dopamine pathway genes NRG1 and LMX1A are associated with cognitive performance in bipolar disorder. *Bipolar Disord.* 2015;17(8):859-68.
<https://doi.org/10.1111/bdi.12347>
3. de Bartolomeis A, Buonaguro EF, Iasevoli F, Tomasetti C. The emerging role of dopamine-glutamate interaction and of the postsynaptic density in bipolar disorder pathophysiology: Implications for treatment. *J Psychopharmacol.* 2014;28(6):505-26.
<https://doi.org/10.1177/0269881114523864>
4. Rowland TA, Marwaha S. Epidemiology and risk factors for bipolar disorder. *Therapeutic Advances in Psychopharmacology.* 2018;8(9):251-69.
<https://doi.org/10.1177/2045125318769235>
5. Tan J, Lin Y, Su L, Yan Y, Chen Q, Jiang H, Wei Q, Gu L. Association between DAOA gene polymorphisms and the risk of schizophrenia, bipolar disorder and depressive disorder. *Prog Neuropsychopharmacol Biol Psychiatry.* 2014;51:89-98.
<https://doi.org/10.1016/j.pnpbp.2014.01.007>
6. Zebarjad F, Saadat M. Association between 15 Insertion/Deletion Polymorphisms and the Risk of Type 1 Bipolar Disorder. *Gene Expression.* 2025;24(2):95-101.
<https://doi.org/10.14218/GE.2023.00159>
7. Oliva V, Fico G, Prisco MD, Gonda X, Rosa AR, Vieta E. Bipolar disorders: an update on critical aspects. *Lancet Reg Health Eur.* 2025;48.
<https://doi.org/10.1016/j.lanepe.2024.101135>
8. Scaini G, Valassori SS, Diaz AP, Lima CN, Benevenuto D, Fries GR, Quevedo J. Neurobiology of bipolar disorders: a review of genetic components, signaling pathways, biochemical changes, and neuroimaging findings. *Braz J Psychiatry.* 2020;42(5):536-51.
<https://doi.org/10.1590/1516-4446-2019-0732>
9. Kalcev G, Preti A, Scano A, Orrù G, Carta MG. Insight into susceptibility genes associated with bipolar disorder: a systematic review. *Eur Rev Med Pharmacol Sci.* 2021;25(18):5701-24.
https://doi.org/10.26355/eurev_202109_26789
10. Maletic V, Raison C. Integrated neurobiology of bipolar disorder. *Front Psychiatry.* 2014;5:98.
<https://doi.org/10.3389/fpsy.2014.00098>
11. Grunze H. Chapter 40 - Bipolar Disorder. In: Zigmund MJ, Rowland LP, Coyle JT, editors. *Neurobiology of Brain Disorders.* San Diego: Academic Press; 2015. p. 655-73.
<https://doi.org/10.1016/B978-0-12-398270-4.00040-9>
12. Newberg AR, Catapano LA, Zarate CA, Manji HK. Neurobiology of bipolar disorder. *Expert Rev Neurother.* 2008;8(1):93-110.
<https://doi.org/10.1586/14737175.8.1.93>
13. Andreazza AC, Young LT. The neurobiology of bipolar disorder: identifying targets for specific agents and synergies for combination treatment. *Int J Neuropsychopharmacol.* 2014;17(7):1039-52.
<https://doi.org/10.1017/S1461145713000096>
14. Sun Z, Ma Y, Li W, He J, Li J, Yang X, Mao P, Cubells JF, Tang YL. Associations between the DBH gene, plasma dopamine β -hydroxylase activity and cognitive measures in Han Chinese patients with schizophrenia. *Schizophr Res.* 2018;193:58-63.
<https://doi.org/10.1016/j.schres.2017.06.028>
15. Gonzalez-Lopez E, Vrana KE. Dopamine beta-hydroxylase and its genetic variants in human health and disease. *J Neurochem.* 2020;152(2):157-81.
<https://doi.org/10.1111/jnc.14893>
16. Sun Z, Bo Q, Mao Z, Li F, He F, Pao C, Li W, He Y, Ma X, Wang C. Reduced Plasma Dopamine- β -Hydroxylase Activity Is Associated With the Severity of Bipolar Disorder: A Pilot Study. *Front Psychiatry.* 2021;12:566091.
<https://doi.org/10.3389/fpsy.2021.566091>
17. Perry SE, Summar ML, Phillips JA, Robertson D. Linkage analysis of the human dopamine beta-hydroxylase gene. *Genomics.* 1991;10(2):493-5.
[https://doi.org/10.1016/0888-7543\(91\)90339-g](https://doi.org/10.1016/0888-7543(91)90339-g)
18. Shakiba M, Hashemi M, Shahrabadi S, Rezaei M, Taheri M. Lack of Association Between Dopamine Beta-Hydroxylase (DBH) 19-bp Insertion/Deletion Polymorphism and Risk of Schizophrenia. *Iran J Psychiatry.* 2016;11(4):239-43.
<https://doi.org/10.1159/000495238>

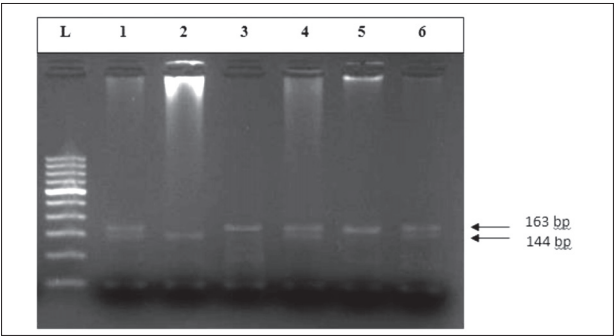
19. Hui L, Zhang X, Huang XF, Han M, Fernandez F, Yu Y, Sun S, Li W, Chen DC, Xiu MH, Kosten TR, Zhang XY. The dopamine b-hydroxylase 19 bp insertion/deletion polymorphism was associated with first-episode but not medicated chronic schizophrenia. *J Psychiatr Res.* 2012;46(6):733-7. <https://doi.org/10.1016/j.jpsychires.2012.02.016>
20. Mustapic M, Maihofer AX, Mahata M, Chen Y, Baker DG, O'Connor DT, Nievergelt CM. The catecholamine biosynthetic enzyme dopamine β -hydroxylase (DBH): first genome-wide search positions trait-determining variants acting additively in the proximal promoter. *Hum Mol Genet.* 2014;23(23):6375-84. <https://doi.org/10.1093/hmg/ddu332>
21. Andreou D, Saetre P, Werge T, Andreassen OA, Agartz I, Sedvall GC, Hall H, Terenius L, Jönsson EG. D-amino acid oxidase activator gene (DAOA) variation affects cerebrospinal fluid homovanillic acid concentrations in healthy Caucasians. *Eur Arch Psychiatry Clin Neurosci.* 2012;262(7):549-56. <https://doi.org/10.1007/s00406-012-0313-z>
22. Ahmadi L, Kazemi Nezhad SR, Behbahani P, Khajeddin N, Pourmehdi-Boroujeni M. Genetic Variations of DAOA (rs947267 and rs3918342) and COMT Genes (rs165599 and rs4680) in Schizophrenia and Bipolar I Disorder. *Basic Clin Neurosci.* 2018;9(6):429-38. <https://doi.org/10.32598/bcn.9.6.429>
23. Young AH, Juruena MF. The Neurobiology of Bipolar Disorder. In: Young AH, Juruena MF, editors. *Bipolar Disorder: From Neuroscience to Treatment*. Cham: SpringerM 2020. p. 179. (Current Topics in Behavioral Neurosciences; Vol 48). https://doi.org/10.1007/7854_2020_179
24. Serretti A, Mandelli L. The genetics of bipolar disorder: genome "hot regions," genes, new potential candidates and future directions. *Mol Psychiatry.* 2008;13(8):742-71. <https://doi.org/10.1038/mp.2008.29>
25. Gatt JM, Burton KLO, Williams LM, Schofield PR. Specific and common genes implicated across major mental disorders: a review of meta-analysis studies. *J Psychiatr Res.* 2015;60:1-13. <https://doi.org/10.1016/j.jpsychires.2014.09.014>
26. Ates O, Celikel FC, Taycan SE, Sezer S, Karakus N. Association between 1603C>T polymorphism of DBH gene and bipolar disorder in a Turkish population. *Gene.* 2013;519(2):356-9. <https://doi.org/10.1016/j.gene.2013.01.031>
27. Boukhenaf Y, Ayachi OS, Achou R, Bernou AI, Madoui FZ, Sifi K, Rezgoun ML. Association of dopamine receptor D2 -141C insertion/deletion and dopamine beta-hydroxylase 19 bp insertion/deletion polymorphisms with schizophrenia: A case-control study in the eastern Algerian population. *Arch Biol Sci.* 2024;76(3):313-24. <https://doi.org/10.2298/ABS240526023B>
28. Smith EN, Bloss CS, Badner JA, Barrett T, Belmonte PL, Berrettini W, Byerley W, Coryell W, Craig D, Edenberg HJ, Eskin E, Foroud T, Gershon E, Greenwood TA, Hipolito M, Koller DL, Lawson WB, Liu C, Lohoff F, McInnis MG, McMahon FJ, Mirel DB, Murray SS, Nievergelt C, Nurnberger J, Nwulia EA, Paschall J, Potash JB, Rice J, Schulze TG, Scheftner W, Panganiban C, Zaitlen N, Zandi PP, Zöllner S, Schork NJ, Kelsoe JR. Genome-wide association study of bipolar disorder in European American and African American individuals. *Mol Psychiatry.* 2009;14(8):755-63. <https://doi.org/10.1038/mp.2009.43>
29. Bass NJ, Datta SR, McQuillin A, Puri V, Choudhury K, Thirumalai S, Lawrence J, Quedsted D, Pimm J, Curtis D, Gurling HMD. Evidence for the association of the DAOA (G72) gene with schizophrenia and bipolar disorder but not for the association of the DAO gene with schizophrenia. *Behav Brain Funct.* 2009;5(1):28. <https://doi.org/10.1186/1744-9081-5-28>
30. Müller DJ, Zai CC, Shinkai T, Strauss J, Kennedy JL. Association between the DAOA/G72 gene and bipolar disorder and meta-analyses in bipolar disorder and schizophrenia. *Bipolar Disorders.* 2011;13(2):198-207. <https://doi.org/10.1111/j.1399-5618.2011.00905.x>
31. Gaysina D, Cohen-Woods S, Chow PC, Martucci L, Schosser A, Ball HA, Tozzi F, Perry J, Muglia P, Kennedy JL, King N, Vincent JB, Parikh SV, Strauss J, Craig IW, McGuffin P, Farmer A. Association analysis of DAOA and DAO in bipolar disorder: results from two independent case-control studies. *Bipolar Disorders.* 2010;12(5):579-81. <https://doi.org/10.1111/j.1399-5618.2010.00837.x>
32. Schumacher J, Jamra RA, Freudenberger J, Becker T, Ohlraun S, Otte ACJ, Tullius M, Kovalenko S, Van Den Bogaert A, Maier W, Rietschel M, Propping P, Nöthen MM, Cichon S. Examination of G72 and D-amino-acid oxidase as genetic risk factors for schizophrenia and bipolar affective disorder. *Mol Psychiatry.* 2004;9(2):203-7. <https://doi.org/10.1038/sj.mp.4001421>
33. American Psychiatric Association. *Diagnostic and statistical manual of mental disorders: DSM-5TM*. 5th ed. Arlington, VA, US: American Psychiatric Publishing, Inc.; 2013. xlv, 947 p. (Diagnostic and statistical manual of mental disorders: DSM-5TM).
34. Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucl Acids Res.* 1988;16(3):1215. <https://doi.org/10.1093/nar/16.3.1215>
35. Kirov G, Jones I, McCandless F, Craddock N, Owen MJ. Family-based association studies of bipolar disorder with candidate genes involved in dopamine neurotransmission: DBH, DAT1, COMT, DRD2, DRD3 and DRD5. *Mol Psychiatry.* 1999;4(6):558-65. <https://doi.org/10.1038/sj.mp.4000565>
36. Hess C, Reif A, Strobel A, Boreatti-Hümmer A, Heine M, Lesch KP, Jacob CP. A functional dopamine-beta-hydroxylase gene promoter polymorphism is associated with impulsive personality styles, but not with affective disorders. *J Neural Transm.* 2009;116(2):121-30. <https://doi.org/10.1007/s00702-008-0138-0>
37. Togsverd M, Werge TM, Tankó LB, Bagger YZ, Hansen T, Qin G, Christiansen C, Rasmussen HB. Association of a dopamine beta-hydroxylase gene variant with depression in elderly women possibly reflecting noradrenergic dysfunction. *J Affect Disord.* 2008;106(1-2):169-72. <https://doi.org/10.1016/j.jad.2007.06.010>
38. Zhou Y, Wang J, He Y, Zhou J, Xi Q, Song X, Ye Y, Ying B. Association between dopamine beta-hydroxylase 19-bp

- insertion/deletion polymorphism and major depressive disorder. *J Mol Neurosci*. 2015;55(2):367–71.
<https://doi.org/10.1007/s12031-014-0339-y>
39. Zhang XY, Chen DC, Xiu MH, Hui L, Liu H, Luo X, Zuo L, Zhang H, Kosten TA, Kosten TR. Association of functional dopamine-beta-hydroxylase (DBH) 19 bp insertion/deletion polymorphism with smoking severity in male schizophrenic smokers. *Schizophrenia Research*. 2012;141(1):48–53. <https://doi.org/10.1016/j.schres.2012.07.011>
 40. Yamamoto K, Cubells JF, Gelernter J, Benkelfat C, Lalonde P, Bloom D, Lal S, Labelle A, Turecki G, Rouleau GA, Joobor R. Dopamine beta-hydroxylase (DBH) gene and schizophrenia phenotypic variability: a genetic association study. *Am J Med Genet B Neuropsychiatr Genet*. 2003;117B(1):33–8.
<https://doi.org/10.1002/ajmg.b.10011>
 41. Tang Y, lang, Epstein MP, Anderson GM, Zabetian CP, Cubells JF. Genotypic and haplotypic associations of the DBH gene with plasma dopamine β -hydroxylase activity in African Americans. *Eur J Hum Genet*. 2007;15(8):878–83.
<https://doi.org/10.1038/sj.ejhg.5201838>
 42. Howe AS, Leung T, Bani-Fatemi A, Souza R, Tampakeras M, Zai C, Kennedy JL, Strauss J, De Luca V. Lack of association between dopamine- β hydroxylase gene and a history of suicide attempt in schizophrenia: comparison of molecular and statistical haplotype analyses. *Psychiatric Genetics*. 2014;24(3):110.
<https://doi.org/10.1097/YPG.0000000000000031>
 43. Cubells JF, Sun X, Li W, Bonsall RW, McGrath JA, Avramopoulos D, Lasseter VK, Wolyniec PS, Tang YL, Mercer K, Pulver AE, Elston RC. Linkage analysis of plasma dopamine β -hydroxylase activity in families of patients with schizophrenia. *Hum Genet*. 2011;130(5):635–43.
<https://doi.org/10.1007/s00439-011-0989-6>
 44. Ji N, Shuai L, Chen Y, Liu L, Li H, Li Z, Yang L, Qian Q, Tang YL, Cubells JF, Wang Y. Dopamine β -hydroxylase gene associates with stroop color-word task performance in Han Chinese children with attention deficit/hyperactivity disorder. *Am J Med Genet B Neuropsychiatr Genet*. 2011;156B(6):730–6. <https://doi.org/10.1002/ajmg.b.31215>
 45. Schulze TG, Ohlraun S, Czerski PM, Schumacher J, Kassem L, Deschner M, Gross M, Tullius M, Heidmann V, Kovalenko S, Abou Jamra R, Becker T, Leszczynska-Rodziewicz A, Hauser J, Illig T, Klopp N, Wellek S, Cichon S, Henn FA, McMahon FJ, Maier W, Propping P, Nöthen MM, Rietschel M. Genotype-phenotype studies in bipolar disorder showing association between the DAOA/G30 locus and persecutory delusions: a first step toward a molecular genetic classification of psychiatric phenotypes. *Am J Psychiatry*. 2005;162(11):2101–8.
<https://doi.org/10.1176/appi.ajp.162.11.2101>
 46. Zhang Z, Li Y, Zhao Q, Huang K, Wang P, Yang P, Li S, Feng G, Lindpaintner K, He L, Shi Y. First evidence of association between G72 and bipolar disorder in the Chinese Han population. *Psychiatr Genet*. 2009;19(3):151–3.
<https://doi.org/10.1097/YPG.0b013e32832a50f1>
 47. Gomez L, Wigg K, Feng Y, Kiss E, Kapornai K, Tamás Z, Mayer L, Baji I, Daroczi G, Benak I, Kothencne VO, Dombovari E, Kaczvinsk E, Besnyo M, Gadoros J, King N, Szekely J, Kovacs M, Vetro A, Kennedy JL, Barr CL. G72/G30 (DAOA) and juvenile-onset mood disorders. *Am J Med Genet B Neuropsychiatr Genet*. 2009;150B(7):1007–12.
<https://doi.org/10.1002/ajmg.b.30904>
 48. Gawlik M, Wehner I, Mende M, Jung S, Pfuhmann B, Knapp M, Stöber G. The DAOA/G30 locus and affective disorders: haplotype based association study in a polydiagnostic approach. *BMC Psychiatry*. 2010;10:59.
<https://doi.org/10.1186/1471-244X-10-59>
 49. Prata D, Breen GD, Osborne S, Munro J, St Clair DM, Collier D. Association of DAO and G72(DAOA)/G30 genes with bipolar affective disorder. *Am J Med Genet B Neuropsychiatr Genet*. 2008;147B(6):914–7.
<https://doi.org/10.1002/ajmg.b.30682>
 50. Chumakov I, Blumenfeld M, Guerassimenko O, Cavarec L, Palicio M, Abderrahim H, Bougueleret L, Barry C, Tanaka H, La Rosa P, Puech A, Tahri N, Cohen-Akenine A, Delabrosse S, Lissarrague S, Picard FP, Maurice K, Essieux L, Millasseau P, Grel P, Debailleul V, Simon AM, Caterina D, Dufau I, Malekzadeh K, Belova M, Luan JJ, Bouillot M, Sambucy JL, Primas G, Saumier M, Boubkiri N, Martin-Saumier S, Nasroune M, Peixoto H, Delaye A, Pinchot V, Bastucci M, Guillou S, Chevillon M, Sainz-Fuertes R, Meguenni S, Aurich-Costa J, Cherif D, Gimalac A, Van Duijn C, Gauvreau D, Ouellette G, Fortier I, Raelson J, Sherbatich T, Riazanskaia N, Rogaev E, Raeymaekers P, Aerssens J, Konings F, Luyten W, Maciardi F, Sham PC, Straub RE, Weinberger DR, Cohen N, Cohen D. Genetic and physiological data implicating the new human gene G72 and the gene for D-amino acid oxidase in schizophrenia. *Proc Natl Acad Sci U S A*. 2002;99(21):13675–80.
<https://doi.org/10.1073/pnas.182412499>
 51. Grigoriu-Serbanescu M, Herms S, Diaconu CC, Jamra RA, Meier S, Bleotu C, Neagu AI, Prelipceanu D, Sima D, Gherghel M, Mihailescu R, Rietschel M, Nöthen MM, Cichon S, Mühleisen TW. Possible association of different G72/G30 SNPs with mood episodes and persecutory delusions in bipolar I Romanian patients. *Prog Neuropsychopharmacol Biol Psychiatry*. 2010;34(4):657–63.
<https://doi.org/10.1016/j.pnpbp.2010.03.008>
 52. Alves V de M, Pereira e Silva AC, Neto VL de M, Andrade TG, Nardi AE. Associations between genetic polymorphisms and bipolar disorder. *Rev Psiq Clin*. 2012;39(1):34–9.
 53. Sigitova E, Fišar Z, Hroudová J, Cikánková T, Raboch J. Biological hypotheses and biomarkers of bipolar disorder. *Psychiatry Clin Neurosci*. 2017;71(2):77–103.
<https://doi.org/10.1111/pcn.12476>
 54. Gigante AD, Bond DJ, Lafer B, Lam RW, Young LT, Yatham LN. Brain glutamate levels measured by magnetic resonance spectroscopy in patients with bipolar disorder: a meta-analysis. *Bipolar Disord*. 2012;14(5):478–87.
<https://doi.org/10.1111/j.1399-5618.2012.01033.x>
 55. Zuliani R, Moorhead TWJ, Job D, McKirdy J, Sussmann JED, Johnstone EC, Johnstone EC, Lawrie SM, Brambilla P, Hall J, McIntosh AM. Genetic variation in the G72 (DAOA) gene affects temporal lobe and amygdala structure in subjects affected by bipolar disorder. *Bipolar Disord*. 2009;11(6):621–7.
<https://doi.org/10.1111/j.1399-5618.2009.00731.x>

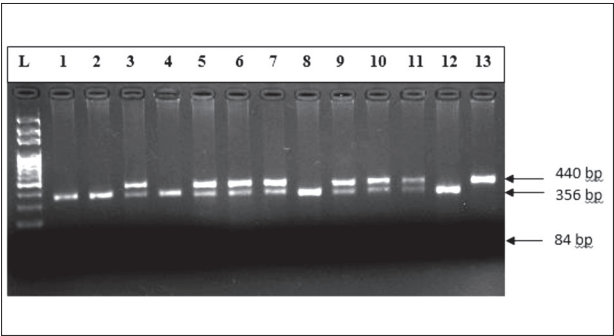
56. Pauli A, Prata DP, Mechelli A, Picchioni M, Fu CHY, Chaddock CA, Kane F, Kalidindi S, McDonald C, Kravariti E, Touloupoulou T, Bramon E, Walshe M, Ehler N, Georgiades A, Murray R, Collier DA, McGuire P. Interaction between

effects of genes coding for dopamine and glutamate transmission on striatal and parahippocampal function. *Hum Brain Mapp.* 2013;34(9):2244–58. <https://doi.org/10.1002/hbm.22061>

SUPPLEMENTARY MATERIALS



Supplementary Fig. S1. Electrophoresis profile of *DBH* rs72393728 by PCR. Lane L – ladder (50 bp); lanes 1, 4, and 6 are heterozygotes (InsDel); lane 2 is homozygote (DelDel); lanes 3 and 5 are homozygotes (InsIns).



Supplementary Fig. S2. Electrophoresis profile of *DAOA* rs3918342 by PCR-RFLP. Lane L – ladder 100 bp; lanes 1, 2, 4, 8, and 12 are homozygotes (CC); lanes 3, 5, 6, 7, 9, 10, and 11 are heterozygotes (CT); lane 13 is homozygote (TT).

ONLINE SUPPLEMENTARY RESEARCH DATASET

The raw data underlying this article is available as an online supplementary research dataset: https://www.serbiosoc.org.rs/NewUploads/Uploads/Bernou%20et%20al_11120-Dataset.pdf