

## The role of potassium channels and calcium in the relaxation mechanism of magnesium sulfate on the isolated rat uterus

Dragana Sokolović<sup>1</sup>, Dragana Drakul<sup>1</sup>, Zorana Oreščanin Dušić<sup>2</sup>, Nikola Tatalović<sup>2</sup>, Milica Pecelj<sup>3,4,5</sup>, Slobodan Milovanović<sup>1</sup> and Duško Blagojević<sup>2,\*</sup>

<sup>1</sup> Department of Pharmacology, Faculty of Medicine at Foča, University of East Sarajevo, East Sarajevo, 73301 Foča, Republic of Srpska, Bosnia and Herzegovina

<sup>2</sup> Department for Physiology, Institute for Biological Research "Siniša Stanković", University of Belgrade, 11000 Belgrade, Serbia

<sup>3</sup> Faculty of Philosophy, University of East Sarajevo, Pale, Republic of Srpska, Bosnia and Herzegovina

<sup>4</sup> Geographical Institute "Jovan Cvijić" of the Serbian Academy of Sciences and Arts, 11000 Belgrade, Serbia

<sup>5</sup> Institute of Sports, Tourism and Service, South Ural State University, Chelyabinsk, Russia

\*Corresponding author: [dblagoje@ibiss.bg.ac.rs](mailto:dblagoje@ibiss.bg.ac.rs)

Received: June 15, 2018; Revised: July 6, 2018; Accepted: July 6, 2018; Published online: July 16, 2018

**Abstract:** MgSO<sub>4</sub> is used as a tocolytic agent. It is considered to be a calcium channel antagonist, but a different mechanism of its action might be involved. The aim of this study was to examine the contribution of calcium concentrations and potassium channels in the mechanism of MgSO<sub>4</sub>-mediated uterine relaxation. Isolated uteri from female Wistar rats were treated with increasing MgSO<sub>4</sub> concentrations (0.1-30 mM). MgSO<sub>4</sub> induced dose-dependent inhibition of spontaneous activity. Addition of Ca<sup>2+</sup> (6 mM and 12 mM) stimulated uterine contractile activity and attenuated the inhibitory activity of MgSO<sub>4</sub>. In order to analyze the role of different subtypes of potassium channels, Ca<sup>2+</sup>-stimulated uteri were pretreated with glibenclamide (Glib), a selective ATP-sensitive potassium channel inhibitor (K<sub>ATP</sub>), tetraethylammonium (TEA), a non-specific inhibitor of large conductance calcium-activated potassium channels (BK<sub>Ca</sub>), and 4-aminopyridine (4-AP), a voltage-sensitive potassium channel inhibitor (K<sub>v</sub>), at concentrations that had no effect *per se*. Pretreatment with 4-AP had no effect on MgSO<sub>4</sub>-mediated relaxation of Ca<sup>2+</sup>-stimulated uteri. The relaxing effect of MgSO<sub>4</sub> was potentiated by pretreatment with glibenclamide. Pretreatment with TEA attenuated the MgSO<sub>4</sub>-mediated decrease in frequency. Our results suggest that MgSO<sub>4</sub> acts as a general calcium antagonist that influences Ca<sup>2+</sup>-mediated potassium channels. Furthermore, it seems that MgSO<sub>4</sub> uterine relaxation activity is partially mediated by selective ATP-sensitive potassium channels, suggesting an ATP-dependent role.

**Keywords:** MgSO<sub>4</sub>; uterus; K<sup>+</sup> channels; Ca<sup>2+</sup> channels; tocolytic

### INTRODUCTION

Magnesium sulfate (MgSO<sub>4</sub>, mineral salt, soluble in water) is used as a laxative, tocolytic agent and it is known as a functional blocker of calcium channels [1,2]. Despite the long-standing experience of its application, the use of MgSO<sub>4</sub> in gynecology has been a source of controversy for years. MgSO<sub>4</sub> was first used in 1906 to prevent eclamptic attacks by Horn in Germany, when administered intrathecally [3]. Its intramuscular use was first performed in 1926 to prevent repeated attacks in women with eclampsia [4], while the first intravenous administration was in 1933 to women with eclampsia and preeclampsia [5].

The tocolytic effects of MgSO<sub>4</sub> were originally described by Hall et al. in 1959 [6]. Stallworth et al. (1981) found a slight decrease in the incidence of uterine contractions, but no significant change in the intensity of contractions during MgSO<sub>4</sub> administration [7]. Meta-analysis has shown that that magnesium reduces the risk of birth within 48 h by 15%, but it is considered not significant [8]. However, a combination of a betamimetic agonist and MgSO<sub>4</sub> has been introduced, and studies have shown its efficiency in prolonging gestation [9-11]. On the other hand, tocolytic efficacy was not improved and side effects were increased [12]. Although there are many studies

dealing with the tocolytic effect of  $\text{MgSO}_4$ , the exact mechanism of its action is still unknown.

Different types of  $\beta$ 2-adrenergic agonists,  $\text{Ca}^{2+}$  channel blocker, oxytocin receptor antagonist and nonsteroidal antiinflammatory drugs are also used as tocolytics, but their insufficient effectiveness and side effects compromise their preliminary use [13,14]. Therefore, agents with potential tocolytic characteristics are still needed, and they could include calcium antagonists, potassium channel openers and other vasodilators [15]. Potassium channels are abundant and active in the smooth muscle of the uterus [16-18]. Based on structure and function, the channels are categorized in different groups ( $\text{K}_v$  channels,  $\text{BK}_{\text{Ca}}$  channels, ATP-sensitive potassium channels), and each group contains many subtypes and isoforms [19]. Large conductance calcium-activated potassium channels ( $\text{BK}_{\text{Ca}}$ ) are dominant and active in uterine smooth muscles, especially during gestation [20]. ATP-dependent potassium channels ( $\text{K}_{\text{ATP}}$ ) in the smooth muscles of the uterus form the connection between the metabolic state of the cell and uterine contractility [20,21,22]. Therefore, studying the influence of potassium channel modulators on uterine tissue has been suggested as important for finding new therapeutic concepts in the treatment of uterine contractility disturbances [15,23].

Although wide-scale examinations of drugs as modulators of contractility have been performed, a final therapeutic preference has been omitted [17,23,24].  $\text{MgSO}_4$  is considered a general calcium antagonist, but the potential site of  $\text{MgSO}_4$  cellular physiological activity can also be at the level of potassium channels. Potassium channels are widespread in all living cells and very important for regulating cell membrane excitability [25]. Therefore, the aim of our study was to explore the effect of  $\text{MgSO}_4$  on uterine contractility with regard to the role of calcium concentrations and potassium channels.

## MATERIALS AND METHODS

### Experimental system

All animals were treated according to directive 2010/63/EU of the European Parliament on the protection of animals used for scientific purposes and experiments

were approved by the Ethical Committee for the Use of Laboratory Animals of the Faculty of Medicine Foča, University of East Sarajevo, Decision No. 01-3-88. Animals were kept at 22°C, housed 3 per cage and fed *ad libitum*. Uteri from intact Wistar rats (250-300 g) in the estrus phase of the estrus cycle, determined by examination of a daily vaginal lavage [26], were used.

### Reagents

$\text{MgSO}_4$  was supplied by Galenika a.d. (Belgrade, Serbia). Tetraethylammonium, glibenclamide and 4-aminopyridine were purchased from Sigma-Aldrich (St. Louis, MO, USA). Salts for De Jalon's solution were obtained from Zorka Pharma (Šabac, Serbia), Merck (New Jersey, USA) and Centrohema d.o.o. (Stara Pazova, Serbia).

### Isolated organ bath studies

All rats were killed by rapid decapitation. The uterine horns were rapidly excised, carefully cleaned of surrounding connective tissue and mounted vertically in a 10-mL-volume organ bath containing De Jalon's solution ( $\text{NaCl}$  154 mM,  $\text{KCl}$  5.6 mM,  $\text{CaCl}_2 \times 2\text{H}_2\text{O}$  0.41 mM,  $\text{NaHCO}_3$  5.9 mM and glucose 2.8 mM), under 1 g tension, aerated with 95% oxygen and 5%  $\text{CO}_2$  at 37°C. Experiments were performed after an equilibration period of about 30 min. The effect of  $\text{MgSO}_4$  was examined on a spontaneously active uterus (incubated for 30 min in an organ bath in De Jalon's solution at 37°C, oxygenated with 95% of  $\text{O}_2$  and 5%  $\text{CO}_2$ ), as well as on calcium-stimulated (6 and 12 mM  $\text{Ca}^{2+}$ , the latter was referred to as double Ca) uteri. In order to analyze the possible role of different subtypes of potassium channels,  $\text{Ca}^{2+}$ -stimulated uteri (with 6 mM  $\text{Ca}^{2+}$ ) were pretreated individually with Glib ( $10^{-5}$  M), TEA ( $10^{-3}$  M), or 4-aminopyridine (4-AP,  $10^{-3}$  M). After 10 min, increasing concentrations of  $\text{MgSO}_4$  (0.1-30 mM) were added. Myometrial tension was recorded isometrically with a TSZ-04-E isolated organ bath and transducer (Experimetria, Budapest, Hungary) and an Ugo Basile isolated organ bath and a transducer (Gemonio, Italy).

### Data analysis and statistical procedures

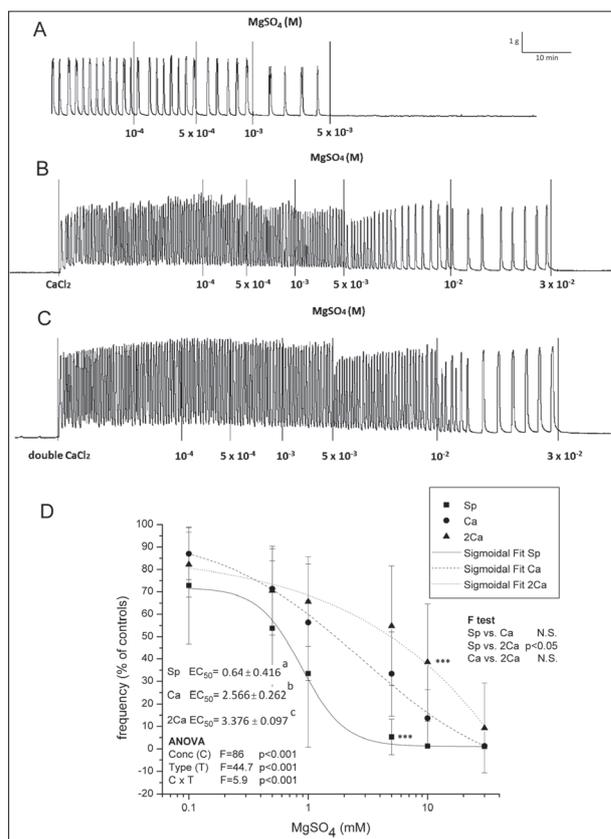
Statistical analyses (descriptive statistics, analysis of variance (ANOVA), *post hoc* tests, F-test and Student's t test) were performed according to the protocols de-

scribed by Hinkle et al. [27] and Manley [28]. Effects of the treatments on uterine contractions were calculated as the percentages of untreated control contractions. Each value is expressed as the mean $\pm$ SD. Differences between groups were analyzed by two-way ANOVA on a logarithmic transformed data row, using concentrations and pretreatments as factors (ANOVA was considered statistically significant when  $p < 0.05$ ), and *post hoc* tested by Tukey's HSD t-test. Since concentration-response curves were sigmoidal in shape, they were fitted according to Boltzmann functions (the concentration axis was logarithmic) and the  $EC_{50}$  were calculated. Sigmoid curves were compared using the F-test.  $EC_{50}$  values were compared using Student's t test (significance:  $p < 0.05$ ).

## RESULTS

MgSO<sub>4</sub> relaxed spontaneous uterine activity in a concentration-dependent manner with regard to frequency (Fig. 1A). There were no significant changes in amplitude until complete cessation of contractions occurred (at the highest used MgSO<sub>4</sub> concentration of 30 mM). The addition of Ca<sup>2+</sup> (6 and 12 mM) caused intensive contractile activity, and these types of uterine activity were referred to as Ca<sup>2+</sup>- or double Ca<sup>2+</sup>-stimulated, according to the concentration of Ca<sup>2+</sup> used. MgSO<sub>4</sub> also relaxed both Ca<sup>2+</sup>- and double Ca<sup>2+</sup>-stimulated active uteri in a concentration-dependent manner (Fig. 1B and C; ANOVA significant concentration effect,  $p < 0.001$ ), but the concentration of MgSO<sub>4</sub> necessary for relaxation of the Ca<sup>2+</sup>- and double Ca<sup>2+</sup>-stimulated active uteri was significantly higher (significant difference in  $EC_{50}$  values, ANOVA significant type and interaction effect,  $p < 0.001$ , significant *post hoc* Tukey t-test; Fig. 1D).  $EC_{50}$  was 5 times higher for double Ca<sup>2+</sup>-stimulated uteri than for the spontaneously active uterus. The addition of both single and double Ca<sup>2+</sup> extended the MgSO<sub>4</sub>-induced relaxation effect and shifted the sigmoid shape for the frequency toward higher concentrations (significant F-test effect,  $p < 0.05$ ; Fig. 1D).

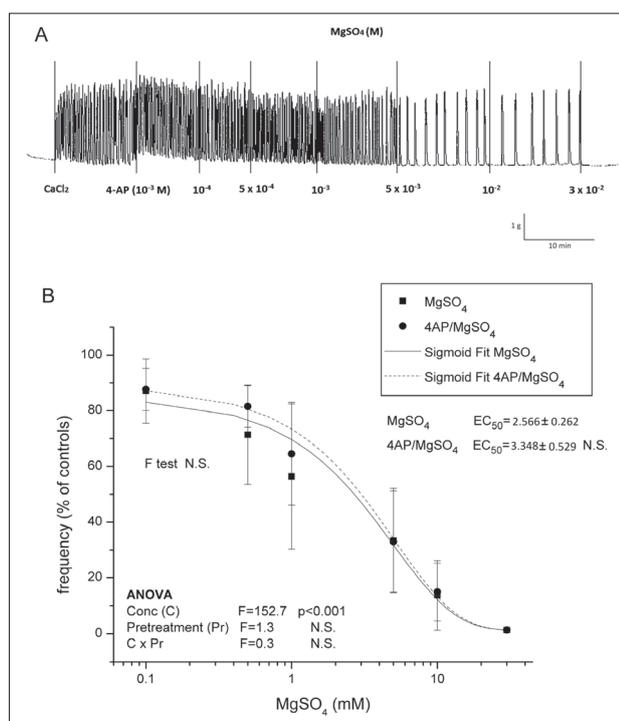
Pretreatment with 4-AP had no effect on MgSO<sub>4</sub>-mediated relaxation of Ca<sup>2+</sup>-stimulated uteri (no significant ANOVA pretreatment effect, no difference between either the sigmoidal fit curve shape or  $EC_{50}$ ; Fig. 2A and B). On the other hand, a relaxing effect



**Fig. 1.** The effect of MgSO<sub>4</sub> on uterine contractile activity under different Ca<sup>2+</sup> concentrations. Graphs show original traces for A – spontaneously active, B – Ca<sup>2+</sup>-stimulated active, and C – double Ca<sup>2+</sup>-stimulated active. Results are presented as the means $\pm$ SD (n=8) for frequency measurements (D). The results were compared by two-way ANOVA for the concentration of MgSO<sub>4</sub> (C) and the type of contractile activity (T) as factors, and F values are presented. From the curves, the  $EC_{50}$  values were calculated, expressed as the mean $\pm$ SD and compared by one-way ANOVA and *post hoc* Tukey's t-test. Differences in the shape of curves were tested by the F-test.

of MgSO<sub>4</sub> was potentiated by pretreatment with glibenclamide (Fig. 3A and B). Pretreatment with glibenclamide significantly deepened relaxation by decreasing frequency (significant ANOVA pretreatment effect,  $p < 0.05$ ). Pretreatment with glibenclamide lowered the concentrations of MgSO<sub>4</sub> that were needed for relaxation (*post hoc* Tukey difference between degrees of relaxation by single equivalent concentration of 5 and 10 mM). There was no difference between the MgSO<sub>4</sub>-induced relaxing effect of Ca<sup>2+</sup>-stimulated and glibenclamide pretreated uteri as regards the amplitude.

Pretreatment with TEA led to the elevation of frequency (significant ANOVA pretreatment effect,

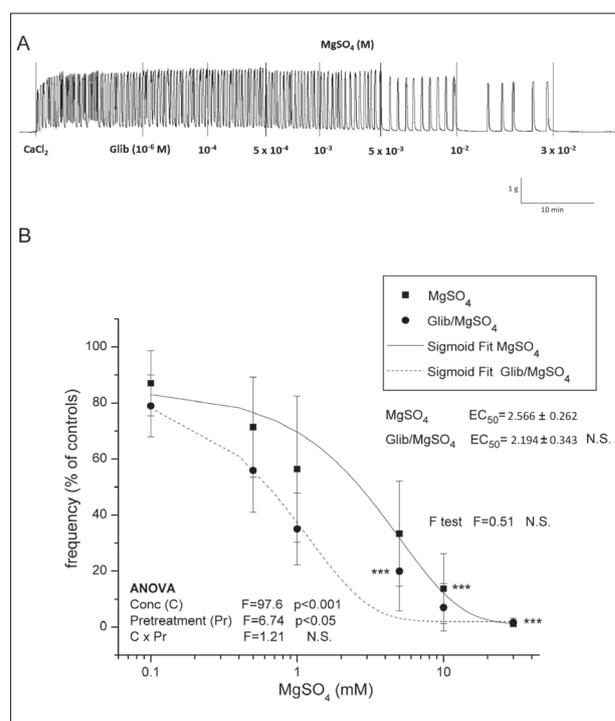


**Fig. 2.** The effect of  $\text{MgSO}_4$  on contractile activity of uteri pretreated with 4-AP. Graphs show original traces for: A –  $\text{Ca}^{2+}$ -stimulated active uteri pretreated with 4-AP. The results are presented as the means $\pm$ SD (n=8) for frequency (B) measurements. The results were compared by two-way ANOVA for the concentration of  $\text{MgSO}_4$  (C) and the type of contractile activity (T) as factors, and F values are presented. From the curves,  $\text{EC}_{50}$  values were calculated, expressed as the mean $\pm$ SD and compared by one-way ANOVA and *post hoc* Tukey's t-test. Differences in the shape of curves were tested by the F-test.

$p < 0.001$ ; Fig. 4A and 4B) after application of  $\text{MgSO}_4$  concentrations above 1 mM (*post hoc* Tukey t test,  $p < 0.001$ ). However, there was no statistically significant difference between  $\text{EC}_{50}$  values for frequency since there were large distributions of data in the middle part of the TEA curve and therefore the SD was high. There was no difference between the  $\text{MgSO}_4$ -relaxing effect of  $\text{Ca}^{2+}$ -stimulated and TEA pretreated uteri regarding amplitude.

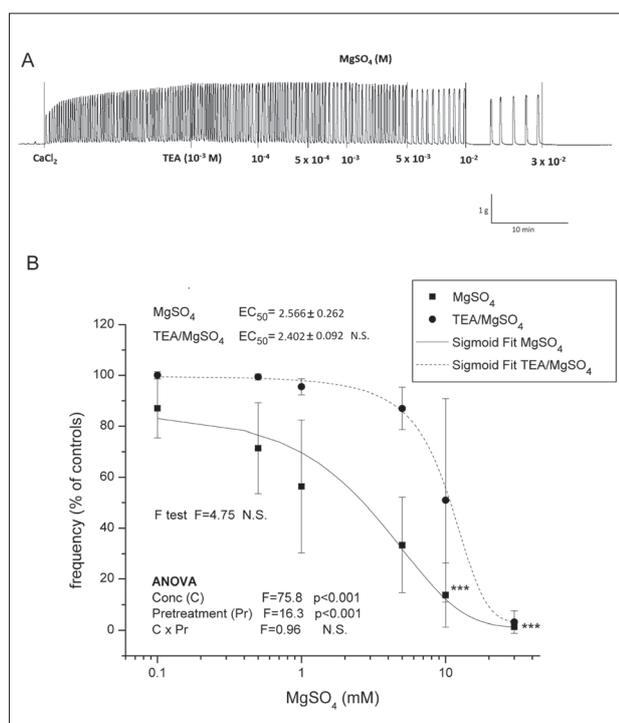
## DISCUSSION

Our results showed that  $\text{MgSO}_4$  inhibited spontaneous uterine activity in a concentration-dependent manner, and that therefore it could be considered as a uterine relaxant. This effect was in the range from 1-30 mM in the *ex vivo* extra uterine fluid (the  $\text{EC}_{50}$  for frequency



**Fig. 3.** The effect of  $\text{MgSO}_4$  on contractile activity of uteri pretreated with glib. Graphs show original traces for: A –  $\text{Ca}^{2+}$ -stimulated active uteri pretreated with Glib. The results are presented as the means $\pm$ SD (n=8) for frequency (B) measurements. The results were compared by two-way ANOVA for the concentration of  $\text{MgSO}_4$  (C) and the type of contractile activity (T) as factors, and F values are presented. From the curves,  $\text{EC}_{50}$  values were calculated, expressed as the mean $\pm$ SD and compared by one-way ANOVA and *post hoc* Tukey's t-test. Differences in the shape of curves were tested by the F-test.

was 2.6 mM). These external  $\text{MgSO}_4$  concentrations are not toxic, but pharmacologically they are selective with regard to the dosage, i.e. the therapeutic window is rather narrow. Magnesium ion concentrations in the plasma and extracellular fluid are approximately 1.2-1.4 mM. One-third is bound by albumin or other proteins and biochemical moieties [29]. This means that small increases in  $\text{Mg}^{2+}$  in the extra uterine fluid can slow down the spontaneous frequency, but for complete relaxation a 3-fold higher concentration is required. Literature data indicate that increasing human serum  $\text{Mg}^{2+}$  concentration by 4-6 mEq/L (2-3 mmol/L) decreases uterine activity in preterm labor [30]. However, our results with pretreatment with glibenclamide and TEA indicated that the effective  $\text{MgSO}_4$  concentration range might be under the influence of applied drugs that can shift its therapeutic potential.



**Fig. 4.** The effect of  $\text{MgSO}_4$  on contractile activity of uteri pretreated with TEA. Graphs show original traces for: **A** –  $\text{Ca}^{2+}$ -stimulated active uteri pretreated with TEA. The results are presented as the means $\pm$ SD ( $n=8$ ) for frequency (**B**) measurements. The results were compared by two-way ANOVA for the concentration of  $\text{MgSO}_4$  (**C**) and the type of contractile activity (T) as factors, and F values are presented. From the curves,  $\text{EC}_{50}$  values were calculated, expressed as the mean $\pm$ SD and compared by one-way ANOVA and *post hoc* Tukey's t-test. Differences in the shape of curves were tested by the F-test.

Our study showed that the addition of  $\text{Ca}^{2+}$  to the isolated organ bath prior to  $\text{MgSO}_4$  significantly attenuated the relaxing effect of  $\text{MgSO}_4$ . Addition of the single  $\text{Ca}^{2+}$  concentration provoked an increase of the force of spontaneous contractions by elevating both amplitude and frequency. This increase was additionally elevated by the double concentration  $\text{Ca}^{2+}$ . In both cases,  $\text{MgSO}_4$  inhibited contractile activity, suggesting its physiological role as a general  $\text{Ca}^{2+}$  antagonist. It is known that  $\text{Mg}^{2+}$  inhibits the ryanodine receptor (RyR)  $\text{Ca}^{2+}$ -release channels by competing with  $\text{Ca}^{2+}$  at the cytosolic activation sites of the channel in the mM range [31-33] and influences the fidelity of coupling between L-type  $\text{Ca}^{2+}$  channels and RyRs [34]. On the other hand, cytosolic levels of  $\text{H}^+$ ,  $\text{Ca}^{2+}$ , adenine nucleotides and  $\text{Mg}^{2+}$  during fatigue influence the gating properties of the SR  $\text{Ca}^{2+}$  channel [35], and the functional roles of

the three main intracellular ions,  $\text{Na}^+$ ,  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$ , are modulated by calmodulin connected voltage-gated  $\text{Na}^+$  channels [36].

Since increasing concentrations of  $\text{Ca}^{2+}$  inhibited the relaxing effect of  $\text{MgSO}_4$  only in part, this indicated the existence of additional cellular signaling pathways that  $\text{Mg}^{2+}$  could operate through. Therefore, we partially inhibited potassium channels by different inhibitors prior to the addition of  $\text{MgSO}_4$ . Our results showed that the voltage-gated  $\text{K}_V$  subfamilies of potassium channels  $\text{K}_V1$ - $\text{K}_V4$  were not involved in the inhibitory action of  $\text{Mg}^{2+}$ , since pretreatment with 4-AP had no effect on concentration-dependent  $\text{Mg}^{2+}$ -promoted uterine relaxation. On the other hand, pretreatment with TEA modified the relaxing activity of  $\text{MgSO}_4$ . TEA is a potent inhibitor of voltage-gated  $\text{K}_V1$ - $\text{K}_V4$  subfamilies of potassium channels, but it also inhibits  $\text{K}_V7$  (KCQN) as well as  $\text{BK}_{\text{Ca}}$  potassium channel subfamilies, suggesting that KCQN as well as  $\text{BK}_{\text{Ca}}$  channels operated during the  $\text{MgSO}_4$ -induced relaxant effect. In our experiment, pretreatment with TEA attenuated the relaxing effect as regards the frequency. Given that XE991, a KCNQ channel inhibitor, elevated the frequency of the murine myometrium [37], it seems that the partial blockade of the potassium channels' pore by TEA in our experiment contributed to the elevated frequency, and that  $\text{MgSO}_4$  operated as a mild KCQN channel inhibitor. It is known that intracellular  $\text{Mg}^{2+}$  enhances the function of  $\text{BK}_{\text{Ca}}$  potassium channels [38] through distinct binding sites and the activation is not directly affected either by voltage or  $\text{Ca}^{2+}$ . However, TEA is also an efficient blocker of this type of channel, and its suppression contributed to other mechanisms of  $\text{Mg}^{2+}$ -induced inhibition of uterine contractility. Moreover,  $\text{BK}_{\text{Ca}}$  channels are in neuronal cells colocalized with voltage-dependent  $\text{Ca}^{2+}$  channels [39,40,41] or RYR [42], and these functional couples seem to be responsive to the  $\text{Ca}^{2+}$  entering into the cytosol and to the control  $\text{Ca}^{2+}$  concentration [40,42,43]. Shi and Cui [38] demonstrated that the competitive inhibition of  $\text{Ca}^{2+}$ -dependent activation of  $\text{BK}_{\text{Ca}}$  channels by  $\text{Mg}^{2+}$  results in a significant reduction of the  $\text{Mg}^{2+}$ -dependent activation at  $[\text{Ca}^{2+}]_i$  of  $\sim 0.1$ - $100 \mu\text{M}$ . Taken together, our results suggest that the overall interplay between  $\text{Ca}^{2+}$  and  $\text{Mg}^{2+}$  is concentration-dependent. Blocking  $\text{BK}_{\text{Ca}}$  requires more  $\text{Ca}^{2+}$  that is prevented by the presence of  $\text{Mg}^{2+}$ . On the other hand, if the  $\text{Ca}^{2+}$  concentration is

in physiological balance with  $Mg^{2+}$ , after the addition of  $Mg^{2+}$ , its  $Ca^{2+}$  antagonist role is potentiated.

Magnesium interacts with organic phosphates such as ATP and PIP2. Intracellular  $Mg^{2+}$  depresses KCNQ currents by binding to PIP2 electrostatically, thus reducing the availability of PIP2 for direct interaction with the channels [44]. Most of cytoplasmic  $Mg^{2+}$  is in the form of a complex with ATP, phosphonucleotides and phosphometabolites and  $Mg^{2+}$ , with ATP constituting the largest metabolic pool capable of binding  $Mg^{2+}$  within the cytoplasm and the mitochondrial matrix as well [45,46]. Our results showed that a partial blockade of  $K_{ATP}$  channels by glibenclamide stimulated the relaxing activity of  $MgSO_4$ , which points to its ATP-dependent role.

The results presented in our study show that  $MgSO_4$  acts only partially as a general calcium antagonist. Moreover, a part of its physiological pathway is through  $BK_{Ca}$  channels since the blocking of  $BK_{Ca}$  channels with TEA led to the stimulation of frequency. Since relaxing activity was predominantly achieved by the reduction of frequency, it seems that  $MgSO_4$  is a direct  $K^+$  channel inhibitor, but it also affects  $Ca^{2+}$  availability. Furthermore, the  $MgSO_4$  uterine relaxing activity is influenced by selective ATP-sensitive potassium channels suggesting also an ATP-dependent role.

**Funding:** This work was funded by the Ministry of Education, Science and Technological Development, Republic of Serbia, Grant No. 173014 "Molecular Mechanisms of Redox Signaling in Homeostasis: Adaptation and Pathology".

**Author contributions:** Duško Blagojević, Slobodan Milovanović conceptualized and defined the research idea and created the research design; Zorana Oreščanin-Dušić and Nikola Tatalović searched the literature data; Duško Blagojević selected the statistical tests; Dragana Jokanović, Dragana Drakul, Milica Pecelj, Zorana Oreščanin-Dušić collected and prepared the experimental data; Nikola Tatalović and Duško Blagojević performed the statistical analyses; Dragana Drakul, Milica Pecelj and Dragana Jokanović wrote the first draft of the manuscript; Duško Blagojević, Slobodan Milovanović and Zorana Oreščanin-Dušić wrote the second draft of the manuscript; Nikola Tatalović and Duško Blagojević edited the manuscript.

**Conflict of interest disclosure:** The authors declare that there is no conflict of interests.

## REFERENCES

1. Elliott JP. Magnesium sulfate as a tocolytic agent. *Am J Obstet Gynecol.* 1983;147(3):277-84.
2. Gáspár R, Hajagos-Tóth J. Calcium Channel Blockers as Tocolytics: Principles of Their Actions, Adverse Effects and Therapeutic Combinations. *Pharmaceuticals.* 2013;6:689-99.
3. Chesley LC. History and epidemiology of preeclampsia-eclampsia. *Clinical Obstetrics and Gynecology.* 1984;27(4):801-20.
4. Dorsett L. The intramuscular injection of magnesium sulphate for the control of convulsions in eclampsia. *Am J Obstet Gynecol.* 1926;11:227-31.
5. Lazard EM. An analysis of 575 cases of eclamptic and pre eclamptic toxæmias treated by IV injections of magnesium sulphate. *Am J Obstet Gynecol.* 1933;26:647-56.
6. Hall DG, McGaughey HS, Corey EL, Thornton WN. The effects of magnesium therapy on the duration of labour. *Am J Obstet Gynecol.* 1959;78:27.
7. Stallworth JC, Yeh S, Petrie RH. The effect of magnesium sulphate on fetal heart rate variability and uterine activity. *Am J Obstet Gynecol.* 1981;140:702-6.
8. Crowther CA, Hiller JE, Doyle LW. Magnesium sulphate for preventing preterm birth in threatened preterm labour. *Cochrane Database Syst Rev.* 2002;4:CD001060.
9. Coleman FH. Safety and efficacy of combined ritodrine and magnesium sulfate for preterm labor: a method for reduction of complications. *Am J Perinatol.* 1990;7(4):366-9.
10. Kosasa TS, Busse R, Wahl N, Hirata G, Nakayama RT, Hale RW. Long-term tocolysis with combined intravenous terbutaline and magnesium sulfate: a 10-year study of 1000 patients. *Obstet Gynecol.* 1994;84(3):369-73.
11. Hatjis CG, Nelson LH, Meis PJ, Swain M. Addition of magnesium sulfate improves effectiveness of ritodrine in preventing premature delivery. *Am J Obstet Gynecol.* 1984;150(2):142-50.
12. Ferguson JE, Hensleigh PA, Kredenster D. Adjunctive use of magnesium sulfate with ritodrine for preterm labor tocolysis. *Am J Obstet Gynecol.* 1984;148(2):166-71.
13. Kalezić I, Rodić V, Kitanović S, Milovanović G, Zgradić I, Milovanović S. The effects of ritodrine, on receptors in smooth uterine muscle and heart atria of rats. *Arch Toxicol Kinet Xenobiot Metab.* 1993;1:112-8.
14. van Vliet E, Dijkema GH, Schuit E, Heida KY, Roos C, van der Post J, Parry EC, McCowan L, Lyell DJ, El-Sayed YY, Carr DB, Clark AL, Mahdy ZA, Uma M, Sayin NC, Varol GE, Mol BW, Oudijk MA. Nifedipine maintenance tocolysis and perinatal outcome: an individual participant data meta-analysis. *BJOG.* 2016;123(11):1753-60.
15. Novaković R, Milovanović SR, Heinle H, Protić D, Gojković-Bukarica Lj. The effect of potassium channel opener pinacidil on non-pregnant rat uterus. *Basic Clin Pharmacol Toxicol.* 2007;1742-84.
16. Oreščanin-Dušić Z, Milovanović S, Blagojević D, Nikolić-Kokić A, Radojičić R, Spasojević I, Spasić BM. Diethyldithiocarbamate potentiates the effects of protamine sulfate in the isolated rat uterus. *Redox Rep.* 2009;14:48-54.
17. Oreščanin-Dušić Z, Milovanović S, Radojičić R, Nikolić-Kokić A, Appiah I, Slavić M, Čutura N, Trbojević S, Spasić

- M, Blagojević D. Effects of protamine sulfate on spontaneous and Ca-induced contractile activity in the rat uterus are potassium channels mediated. *Gen. Physiol. Biophys.* 2009;28:143-8.
18. Milovanovic S, Kordic-Bojinović J, Djordjevic S, Drakul D, Sokolovic D, Miletic N, Blagojevic D. The importance of potassium channels in the relaxing effect of pentoxifylline on the isolated rat uterus. *Serb J Exp Clin Res.* 2013;14(2):55-64.
  19. Kuang Q, Purhonen P, Hebert H. Structure of potassium channels. *Cell Mol Life Sci.* 2015;72:3677-93.
  20. Khan RN, Matharoo BB, Arulkumaran S, Ashford ML. Potassium channels in the human myometrium. *Exp Physiol.* 2001;86:255-64.
  21. Morrison JJ, Ashford MLJ, Khan RN, Smith S K. The effects of potassium channel openers on isolated pregnant human myometrium before and after the onset of labor: potential for tocolysis. *Am J Obstet Gynecol.* 1993;169:1277-85.
  22. Appiah I, Nikolic-Kokic A, Orescanin-Dusic Z, Radojčić R, Milovanovic S, Spasic M, Blagojevic D. Reversible Oxidation of Myometrial Voltage-Gated Potassium Channels with Hydrogen Peroxide. *Oxid Med Cell Longev.* 2012;2012:105820.
  23. Kordić-Bojinović J, Oreščanin-Dušić Z, Slavić M, Radojčić R, Spasić M, Milovanović SR, Blagojević D. Effect of indometacin pretreatment on protamine sulfate-mediated relaxation of the isolated rat uterus: the role of the antioxidative defense system. *Pharmacol Rep.* 2011;63(4):1019-28.
  24. Kordić-Bojinović J, Jokanović D, Stanković D, Janković S, Milovanović S. Influence of modulators of relaxant effect of pentoxifylline in isolated rat uterus. *Serb J Exp Clin Res.* 2010;11(3):99-104.
  25. Furchgott RF. Pharmacological characterization of receptors: its relation to radioligand-binding studies. *Fed Proc.* 1978;37(2):115-20.
  26. Marcondes FK, Bianchi FI, Tanno AP. Determination of the estrous cycle phases of rats: some helpful considerations. *Braz J Biol.* 2002;62:609-14.
  27. Hinkle ED, Wiersma W, Jurs GS. Applied statistics for behavioral sciences. 2<sup>nd</sup> ed. Boston: Houghton Mifflin Company; 1994.
  28. Manley BFJ. Multivariate statistical methods. 2<sup>nd</sup> ed. London: Chapman & Hall; 1986.
  29. Romani AMP. Cellular magnesium homeostasis. *Arch Biochem Biophys.* 2011;512(1):1-23.
  30. Guyton ST, Morey TE. Magnesium. In: Atlee JL, editor. Complications in anesthesia. Philadelphia: Saunders, Elsevier; 2007. p. 59-61.
  31. Zahradnikova A, Palade P. Procaine effects on single sarcoplasmic reticulum Ca<sup>2+</sup> release channels. *Biophys J.* 1993;64:991-1003.
  32. Györke I, Györke S. Regulation of the cardiac ryanodine receptor channel by luminal Ca<sup>2+</sup> involves luminal Ca<sup>2+</sup> sensing sites. *Biophys J.* 1998;75:2801-2810.
  33. Laver DR, Baynes TM, Dulhunty AF. Magnesium inhibition of ryanodine-receptor calcium channels: evidence for two independent mechanisms. *J Membr Biol.* 1997;156:213-229.
  34. Zahradníková A, Dura M, Györke I, Escobar A.L, Zahradník I, Györke S. Regulation of dynamic behavior of cardiac ryanodine receptor by Mg<sup>2+</sup> under simulated physiological conditions. *Am J Physiol Cell Physiol* 2003;285:C1059-70.
  35. Coronado R, Morrissette J, Sukhareva M, Vaughan DM. Structure and function of ryanodine receptors. *Am J Physiol Cell Physiol.* 1994;266:C1485-504.
  36. Guo F, Zhou PD, Gao QH, Gong J, Feng R, Xu XX, Liu SY, Hu HY, Zhao MM, Adam HC, Cai JQ, Hao LY. Low-Mg<sup>2+</sup> treatment increases sensitivity of voltage-gated Na<sup>+</sup> channels to Ca<sup>2+</sup>/calmodulin-mediated modulation in cultured hippocampal neurons. *Am J Physiol Cell Physiol.* 2015;308:C594-605.
  37. McCallum LA, Greenwood IA, Tribe RM. Expression and function of Kv7 channels in murine myometrium throughout oestrous cycle. *Pflugers Arch.* 2009;457:1111-20.
  38. Shi J, Cui J. Intracellular Mg<sup>2+</sup> enhances the function of BK-type Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *J Gen Physiol.* 2001;118:589-605.
  39. Yazejian B, DiGregorio DA, Vergara JL, Poage RE, Meriney SD, Grinnell AD. Direct measurements of presynaptic calcium and calcium-activated potassium currents regulating neurotransmitter release at cultured Xenopus nerve-muscle synapses. *J Neurosci.* 1997;17(9):2990-3001.
  40. Yazejian B, Sun XP, Grinnell AD. Tracking presynaptic Ca<sup>2+</sup> dynamics during neurotransmitter release with Ca<sup>2+</sup>-activated K<sup>+</sup> channels. *Nat Neurosci.* 2000;3(6):566-71.
  41. Marrion NV, Tavalin SJ. Selective activation of Ca<sup>2+</sup>-activated K<sup>+</sup> channels by co-localized Ca<sup>2+</sup> channels in hippocampal neurons. *Nature.* 1998;29:900-5.
  42. Jagger JH, Porter VA, Lederer WJ, Nelson MT. Calcium sparks in smooth muscle. *Am J Physiol Cell Physiol.* 2000;278(2):C235-56.
  43. Neher E. Usefulness and limitations of linear approximations to the understanding of Ca<sup>++</sup> signals. *Cell Calcium.* 1998;24(5-6):345-57.
  44. Suh BC, Hille B. Electrostatic interaction of internal Mg<sup>2+</sup> with membrane PIP2 Seen with KCNQ K<sup>+</sup> channels. *J Gen Physiol.* 2007;130(3):241-56.
  45. Scarpa A, Brinley FJ. In situ measurements of free cytosolic magnesium ions. *Fed Proc.* 1981;40(12):2646-52.
  46. Lüthi D, Günzel D, McGuigan JA. Mg-ATP binding: its modification by spermine, the relevance to cytosolic Mg<sup>2+</sup> buffering, changes in the intracellular ionized Mg<sup>2+</sup> concentration and the estimation of Mg<sup>2+</sup> by 31P-NMR. *Exp Physiol.* 1999;84(2):231-52.